

Review Article

An Updated Review of the Molecular Mechanisms in Drug Hypersensitivity

Chun-Bing Chen ^{1,2,3,4,5,6} **Riichiro Abe**,⁷ **Ren-You Pan**,^{1,2} **Chuang-Wei Wang**,^{1,2}
Shuen-Iu Hung ⁸ **Yi-Giien Tsai** ^{9,10,11} and **Wen-Hung Chung** ^{1,2,3,4,6,12}

¹Department of Dermatology, Drug Hypersensitivity Clinical and Research Center, Chang Gung Memorial Hospital, Taipei, Linkou, Keelung, Taiwan

²Chang Gung Immunology Consortium, Chang Gung Memorial Hospital and Chang Gung University, Taoyuan, Taiwan

³Whole-Genome Research Core Laboratory of Human Diseases, Chang Gung Memorial Hospital, Keelung, Taiwan

⁴College of Medicine, Chang Gung University, Taoyuan, Taiwan

⁵Graduate Institute of Clinical Medical Sciences, Chang Gung University, Taoyuan, Taiwan

⁶Immune-Oncology Center of Excellence, Chang Gung Memorial Hospital, Linkou, Taiwan

⁷Division of Dermatology, Niigata University Graduate School of Medical and Dental Sciences, Niigata, Japan

⁸Department and Institute of Pharmacology, School of Medicine, Infection and Immunity Research Center, National Yang-Ming University, Taipei, Taiwan

⁹Division of Pediatric Allergy and Immunology, Changhua Christian Hospital, Changhua City, Taiwan

¹⁰School of Medicine, Kaohsiung Medical University, Kaohsiung, Taiwan

¹¹School of Medicine, Chung Shan Medical University, Taichung, Taiwan

¹²Department of Dermatology, Xiamen Chang Gung Hospital, Xiamen, China

Correspondence should be addressed to Yi-Giien Tsai; 107239@cch.org.tw and Wen-Hung Chung; wenhungchung@yahoo.com

Received 1 September 2017; Accepted 9 November 2017; Published 13 February 2018

Academic Editor: Kurt Blaser

Copyright © 2018 Chun-Bing Chen et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Drug hypersensitivity may manifest ranging from milder skin reactions (e.g., maculopapular exanthema and urticaria) to severe systemic reactions, such as anaphylaxis, drug reactions with eosinophilia and systemic symptoms (DRESS)/drug-induced hypersensitivity syndrome (DIHS), or Stevens–Johnson syndrome (SJS)/toxic epidermal necrolysis (TEN). Current pharmacogenomic studies have made important strides in the prevention of some drug hypersensitivity through the identification of relevant genetic variants, particularly for genes encoding drug-metabolizing enzymes and human leukocyte antigens (HLAs). The associations identified by these studies are usually drug, phenotype, and ethnic specific. The drug presentation models that explain how small drug antigens might interact with HLA and T cell receptor (TCR) molecules in drug hypersensitivity include the hapten theory, the p-i concept, the altered peptide repertoire model, and the altered TCR repertoire model. The broad spectrum of clinical manifestations of drug hypersensitivity involving different drugs, as well as the various pathomechanisms involved, makes the diagnosis and management of it more challenging. This review highlights recent advances in our understanding of the predisposing factors, immune mechanisms, pathogenesis, diagnostic tools, and therapeutic approaches for drug hypersensitivity.

1. Introduction

Drug hypersensitivity reactions are an important public health problem due to their potential to cause life-threatening anaphylaxis and rare severe cutaneous adverse reactions (SCAR). Drug hypersensitivity can be induced

by immunologically mediated reactions (referred as drug allergies) as well as nonallergic direct mast cell-mediated drug reactions. Immunologic reactions have been divided into four categories according to the classical Gell and Coombs system: type I reactions, which are immediate onset and mediated by IgE and mast cells and/or basophils;

type II reactions, which are delayed in onset and caused by antibody- (usually IgG) mediated cell destruction; type III reactions, which are delayed in onset and caused by IgG drug immune complex deposition and complement activation; and type IV reactions, which are delayed in onset and are T cell mediated [1]. According to the World Allergy Organization (WAO), drug hypersensitivity reactions can also be categorized into immediate reactions and delayed reactions based upon the timing of the appearance of symptoms [2].

Immediate-type reactions usually occur within minutes or hours of drug exposure. The clinical manifestations range from pruritus, urticaria, angioedema, and bronchospasm to anaphylaxis. Type I reactions require the presence of drug-specific IgE or the portion of the drug that forms a hapten complex. Drug-specific IgE is produced upon the first exposure to the drug antigen, and then, it binds to basophils or mast cells with the high-affinity Fc receptor. Upon the next exposure to the same drug, two or more IgE molecules on the basophil or mast cell surface may then bind to one multivalent antigen molecule, initiating a series of cellular activation events. This activation causes the extracellular release of granules with preformed inflammatory mediators, including histamine, leukotrienes, prostaglandins, heparin, and other cytokines [3]. IgE-mediated immunologic drug allergy represents a smaller fraction of drug hypersensitivity compared with nonimmunologic drug hypersensitivity [4]. According to the WAO classification system, immunologic anaphylaxis can be caused by an IgE-mediated or non-IgE-mediated mechanism, whereas nonimmunologic anaphylaxis involves direct mast cell activation [2]. Regardless of the underlying mechanism, however, the clinical symptoms of both types of anaphylaxis are similar and often indistinguishable. The mechanism of immediate-type reactions is explained more fully later in this article. In this review, the terminology used to categorize “immediate” or “delayed” drug hypersensitivity is in accordance with the WAO classification system. At the same time, the immediate-type reactions discussed herein are composed of both IgE-mediated reactions as defined by the Gell and Coombs system, as well as non-IgE-mediated and nonimmunologic anaphylactic reactions.

Delayed-type reactions consist primarily of type IV reactions, which are T cell-mediated delayed-type drug hypersensitivity reactions. These reactions usually take several days or even weeks to manifest following drug exposure. These manifestations range from mild maculopapular exanthema (MPE), contact dermatitis, chronic allergic rhinitis, chronic asthma, nephritis, hepatitis, and fixed drug eruptions (FDEs) to life-threatening SCAR. SCAR includes drug reactions with eosinophilia and systemic symptoms (DRESS)/drug-induced hypersensitivity syndrome (DIHS), Stevens–Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN), and acute generalized exanthematous pustulosis (AGEP) [5]. The MPE phenotype consists of self-limited diffuse erythematous macules and papules without systemic involvement [6]. DRESS syndrome, meanwhile, is characterized by cutaneous involvement with typical skin eruptions (e.g., exfoliative dermatitis and generalized maculopapular exanthema), fever, atypical lymphocytosis, eosinophilia,

lymphadenopathy, and systemic involvement (e.g., liver involvement and kidney involvement). This hypersensitivity syndrome was first named after many different terms had already been used to describe the syndrome, with those terms, such as “anticonvulsant hypersensitivity syndrome,” “allopurinol hypersensitivity syndrome,” and “sulfone syndrome,” primarily depending on the culprit drug involved [7, 8]. The term “DRESS” was initially proposed by Bocquet et al. in 1996 in order to provide a more concise description of the syndrome and decrease the ambiguity resulting from the various terms previously used to refer to it [9]. That said, it should be noted that DRESS is also termed “DIHS” by Japanese experts, with the criteria of DRESS as defined by the RegiSCAR group and the criteria of DIHS as defined by Japanese experts being similar, except that HHV-6 reactivation is included in the diagnostic criteria for DIHS [10]. This nosology is somewhat confusing; however, there is a consensus that DRESS and DIHS are likely within the same disease spectrum. Specifically, patients with typical DIHS may represent a severe form of DRESS syndrome [11]. SJS and TEN (SJS/TEN) are characterized as a rapidly progressing blistering exanthema of purpuric macules and target-like lesions accompanied by mucosal involvement and skin detachment. SJS is defined as involving less than 10% body surface area skin detachment, SJS-TEN overlap as involving 10–29%, and TEN as involving more than 30% [12]. AGEP, meanwhile, typically presents as a sudden eruption of small nonfollicular pustules on a background of erythema with systemic involvement along with fever and neutrophilia [13].

Most forms of drug hypersensitivity involve T cell-mediated immune responses against specific drug/peptide antigens, leading to various clinical phenotypes. T cell receptor (TCR), CD4⁺, and CD8⁺ T cells are involved in the different delayed-type drug hypersensitivity reactions [14]. The molecular mechanisms and checkpoints for drug hypersensitivity include T cell activation and immune responses, cytotoxic proteins and cytokine/chemokine secretion, specific TCR clonotypes, impaired drug metabolism or clearance (e.g., the strong association of cytochrome P450 family 2 subfamily C member 9*3 (*CYP2C9*3*) with phenytoin-induced SCAR), and the cell death mechanisms (e.g., miR-18a-5p-induced apoptosis and annexin A1 and formyl peptide receptor 1-induced necroptosis in keratinocytes). In addition, genetic polymorphisms and specific *HLA* loci also play an important role (e.g., *HLA-B*15:02* for carbamazepine- (CBZ-) induced SJS/TEN, *HLA-B*58:01* for allopurinol-induced SCAR, and *HLA-B*57:01* for abacavir-induced hypersensitivity reactions). Moreover, environmental factors, autoimmune disorders, and patients with a prior medical history of viral infection have also been reported to be implicated in susceptibility to drug hypersensitivity.

2. Clinical Perspectives and Variabilities in Severe Drug Hypersensitivity

2.1. Immediate-Type Hypersensitivity. Immediate-type hypersensitivity reactions may range from urticaria and

angioedema to severe fatal reactions, such as bronchospasm and anaphylaxis. Anaphylaxis is a life-threatening systemic hypersensitivity reaction mainly mediated by mast cells and basophil activation via IgE-mediated, non-IgE-mediated, or nonimmunologic mechanisms. Drugs are the most common anaphylaxis triggers in adults, while foods are the most common triggers in children and teenagers [15]. The incidence of drug-induced anaphylaxis has been reported to range from 0.04 to 3.1%, with a mortality rate of around 0.65% [2]. NSAIDs are the main culprits, followed by beta-lactam antibiotics [16, 17]. Perioperative anaphylaxis also remains an issue due to the administration of various combinations of neuromuscular blocking agents (NMBAs), induction agents (e.g., propofol, etomidate, midazolam, and ketamine), and antibiotics [18, 19]. Nonsteroidal anti-inflammatory drugs (NSAIDs) (with the exception of pyrazolones) are believed to rarely be among the causes of IgE-mediated anaphylaxis, but such anaphylaxis is more commonly related to an aberrant arachidonic acid metabolism [20–22]. The non-IgE-mediated immunologic mechanisms can be mediated by IgG antibodies, as well as by complement or contact system activation, but non-IgE-mediated anaphylaxis is clinically indistinguishable from IgE-mediated anaphylaxis [23, 24]. The causes of non-IgE-mediated immunologic anaphylaxis include biologics, lipid incipients, and dextran [2]. In contrast, nonimmunologic anaphylaxis, previously regarded as a form of pseudoallergic drug reaction, involves the direct stimulation of mast cell degranulation. These reactions are limited to certain groups of drugs, including NSAIDs, such as aspirin, as well as opiates, vancomycin, quinolones, and NMBAs [24, 25]. For radiocontrast media-induced anaphylaxis, the mechanisms are not entirely clear and several mechanisms may be involved, including IgE-mediated or direct stimulating histamine release or the activation of the complement cascades [24, 26, 27].

Due to the complexity of NSAID-induced drug hypersensitivity, a panel of experts from the European Academy of Allergy and Clinical Immunology (EAACI) has proposed a classification and practical approach to cases of drug hypersensitivity caused by NSAIDs [28]. The most frequently occurring type of these cases is cross-reactive hypersensitivity, for which the mechanism is not immunological but, rather, is primarily linked to cyclooxygenase-1 inhibition. This immunological type of NSAID-induced hypersensitivity includes NSAID-exacerbated respiratory disease (NERD), NSAID-exacerbated cutaneous disease (NECD), and NSAID-induced urticaria/angioedema (NIUA) [28]. NSAIDs can also induce immunological (noncross-reactive) hypersensitivity reactions, including IgE-mediated single-NSAID-induced urticaria/angioedema or anaphylaxis (SNIUAA), and T cell-mediated single-NSAID-induced delayed hypersensitivity reactions (SNIDHR). Both cross-reactive reactions and SNIUAA are immediate-type reactions [28].

2.2. Delayed-Type Hypersensitivity

2.2.1. Drug Reactions with Eosinophilia and Systemic Symptoms (DRESS)/Drug-Induced Hypersensitivity Syndrome (DIHS). There have been no large epidemiologic studies of

DRESS/DIHS, a shortcoming which could be due to the fact that the term “hypersensitivity syndrome” was instead used before [5]. It could also be explained by the difficulty of diagnosing DRESS/DIHS, which presents with a complex natural course, a wide diversity of manifestations, and various laboratory abnormalities, and also because there is no specific code for this condition [29]. The incidence of anticonvulsant-related DRESS/DIHS is about one per 1000 to one per 10,000 new users [30]. DRESS/DIHS can occur in pediatric patients, but is more common in adults [31]. Antiepileptic agents and allopurinol are the most commonly reported offending medications [32]. The symptoms often begin 2 to 6 weeks after drug incubation [9]. Damage to multiple systemic organs may occur during the course of DRESS/DIHS syndrome. The liver is most commonly involved among the organs, with liver involvement having been found in 51–84% of patients [33, 34]. Renal involvement also occurs frequently, having been reported in 10–57% of patients [33, 34]. Lung involvement is the third most common type of systemic involvement and may present in various forms ranging from nonspecific symptoms to interstitial pneumonitis, pleuritis, and acute respiratory distress syndrome [35, 36]. Cardiac involvement, meanwhile, has been reported in 4–27% of patients with DRESS/DIHS [37]. This complication is likely associated with the fatal outcomes of the condition, especially when acute necrotizing eosinophilic myocarditis occurs [38]. Several other systemic organs can also be involved in DRESS/DIHS, including the gastrointestinal tract, pancreas, central nervous system, and thyroid, while multiple organ failure associated with disseminated intravascular coagulation or hemophagocytic syndrome may also occur [31, 39]. The overall mortality rate of DRESS/DIHS is around 10% [32]. The likelihood of mortality in cases of DRESS/DIHS is primarily determined by the degree of systemic involvement [35]. Tachycardia, leukocytosis, tachypnea, coagulopathy, gastrointestinal bleeding, and systemic inflammatory response syndrome (SIRS) have also been found to be associated with poor outcomes in DRESS/DIHS patients [33].

2.2.2. Stevens-Johnson Syndrome (SJS)/Toxic Epidermal Necrolysis (TEN). Large epidemiologic investigations of SCAR, especially SJS/TEN, have been performed in Europe beginning 30 years [40, 41]. The reported incidence rates of SJS/TEN for various countries and ethnicities have included 0.93–1.89 cases (Germany), 1.2 cases (France (TEN)), 1.4 cases (Italy), 5.76 cases (United Kingdom), 8.0 cases (Han Chinese), and 12.7 cases (United States) per million people per year [5, 40–45]. The large variation among these rates of incidence might be due to differences in the studies reporting them, including differences in the populations studied, generational differences, differing diagnostic criteria, and differing methodologies (such as the use of registration databases or electronic nationwide healthcare databases). SJS/TEN can occur in different age groups, but the incidences of SJS, SJS-TEN, and TEN appear to be lower in US children than in adults [46]. Racial disparities in SJS/TEN incidence were first reported by a large population-based study, which found that SJS/TEN is more strongly associated

with people of nonwhite ethnicities, particularly Asians and blacks [42]. Pharmacogenetic studies, meanwhile, have pointed out that the strength of genetic associations is related to the prevalence with which susceptibility alleles are carried in different ethnic populations, such as *HLA-B*15:02* and *HLA-B*58:01* in Asians [47, 48]. Although the above classical examples partially explain the phenomenon of specific drug hypersensitivity in specific ethnicities with specific genetic factors, not all cases of drug hypersensitivity can be fully elucidated using this approach.

Cases of SJS/TEN are primarily induced by medications, but *Mycoplasma pneumoniae* infection, viral infection, and collagen vascular diseases have also been found to account for a small portion of such cases [49–52]. The European ongoing case-control surveillance of the SCAR (EuroSCAR) group used a case-control study to identify the drugs carrying a high risk of such reactions and found that they included sulfonamides, aromatic convulsants, allopurinol, oxim nonsteroidal anti-inflammatory drugs, and nevirapine [53]. Newly developed drugs, such as anticancer target therapies, also have the potential to induce SJS/TEN [54]. SJS/TEN induced by monoclonal antibodies targeting the coinhibitory immune checkpoint with antiprogrammed death-1 (PD-1) (nivolumab) and anticytotoxic T-lymphocyte-associated protein 4 (CTLA-4) (ipilimumab) has likewise been reported [55, 56]. Proton pump inhibitors, meanwhile, have been known to induce type I hypersensitivity reactions, but they carry some risk of inducing life-threatening type IV hypersensitivity reactions as well [57]. That risk, however, is mostly confined to the first 8 weeks drug exposure, after which the onset of SCAR is much less likely [53]. Meanwhile, the ALDEN (ALgorithm for Drug causality in Epidermal Necrolysis) has been used to provide structured assistance for the assessment of culprit drugs in SJS/TEN patients [58].

The mortality rates of the various forms of SJS/TEN are high, at approximately 10% for SJS, 30% for overlapping SJS/TEN, and 50% for TEN, for an overall rate of about 25% [34, 59]. Indeed, the mortality rate for cases of TEN has remained high, with reported rates of 15.8%–49.0%, even with the overall improvements to health care in recent decades [42, 44, 60]. A disease severity scoring system called SCORTEN (SCORE of Toxic Epidermal Necrolysis) built on seven independent variables (age > 40 years; presence of malignancy; body surface area involved > 10%; serum urea nitrogen level > 28 mg/dL; glucose level > 252 mg/dL; bicarbonate [HCO_3] level < 20 mEq/L; and heart rate > 120 beats per minute) can be used to help predict mortality in individual cases of SJS/TEN [61, 62]. Modified versions of this scoring system may be needed for specific populations, like pediatric patients [63].

2.2.3. Acute Generalized Exanthematous Pustulosis (AGEP).

The annual incidence of AGEP is estimated to be one to five per million [64]. The EuroSCAR group conducted a large case cohort study of 97 validated cases of AGEP [13]. The mean age of the patients was 56 years (range: 4–91 years) [13]. The list of drugs reported to have been involved is extensive, but certain medications such as aminopenicillins, pristinamycin, quinolones, terbinafine, diltiazem,

antimalarials, and Chinese herbs are known to be associated with higher risks of AGEP [13, 65]. The mortality rate of AGEP has been reported to be about 4%, a relatively low rate compared to those of SJS/TEN and DRESS/DIHS [13].

3. Genetic Factors in Drug Hypersensitivity

3.1. Genetic Factors in Immediate-Type Drug Hypersensitivity.

Genetic predisposing factors have been reported in cases of immediate-type drug hypersensitivity resulting from the use of beta-lactams, aspirin, and other NSAIDs. Interestingly, HLA class II genes (*HLA-DRA* and the *HLA-DRA|HLA-DRB5* interregion) have been linked to immediate reactions to beta-lactams (Table 1) [66]. The genetic variants of proinflammatory cytokines (*IL4*, *IL13*, *IL10*, *IL18*, *TNF*, and *IFNGR1*), the cytokine receptor (*IL4R*), the genes involved in the IgE/FcεRI pathway (the galectin-3 gene (*LGALS3*)), and nucleotide-binding oligomerization domain (*NOD*) gene polymorphisms are also strongly associated with beta-lactam-induced immediate reactions (Table 2) [67–73].

The involvements of *HLA-DRA*, *IL4R*, *NOD2*, and *LGALS3* have also been further validated by a replication study [72]. *HLA-DRB1*13:02* and *HLA-DRB1*06:09* are associated, meanwhile, with aspirin-induced urticaria/angioedema [74]. In addition, *HLA-B44* and *HLA-Cw5* have also been reported to be associated with chronic idiopathic urticaria associated with aspirin- and/or NSAID-induced hypersensitivity [75]. Several genetic predisposing factors have been reported to be associated with immediate-type aspirin hypersensitivity, with those factors involving cytokines (*TGFB1*, *TNF*, and *IL18*) and the production and release of mediators (*LTC4S*, *TBXA2R*, *PTGER4*, *FCER1A*, *MS4A2*, *FCER1G*, and *HNMT*) [76, 77]. Immediate-type hypersensitivity to NSAIDs has also been reported to be associated with genes belonging to the arachidonic acid pathway (*ALOX5*, *ALOX5AP*, *ALOX15*, *TBXAS1*, *PTGDR*, and *CYSLTR1*) [72, 78]. However, the association of common genetic variations in histamine receptor genes was not found in patients with hypersensitivity to NSAIDs [79].

3.2. Genetic Factors in Delayed-Type Drug Hypersensitivity.

Recently, the number of pharmacogenetic studies of HLA-associated drug hypersensitivity and related drug-induced syndromes, such as fixed drug reaction, delayed rash, lupus erythematosus, drug-induced liver disease, DRESS/DIHS, SJS, and TEN, has been increasing. These associations are usually drug and ethnic specific (Table 1), which implies that specific HLA molecules may have higher binding affinities for specific drug antigens and present the drug antigens to specific TCRs, causing a series of T cell activations and adverse immune responses.

3.2.1. Aromatic Anticonvulsants.

Aromatic anticonvulsants, such as carbamazepine (CBZ), phenytoin (PHT), oxcarbazepine (OXC), and lamotrigine (LTG), are known to carry higher risks of inducing SCAR. A strong genetic association between *HLA-B*15:02* and CBZ-induced SJS/TEN was found in 2004 in Han Chinese (corrected P value = 3.1×10^{-27} , odds ratio (OR) = 2504, and 95% confidence interval

TABLE 1: HLA association with various phenotypes of drug hypersensitivity in different populations.

Associated drug	HLA allele	Hypersensitivity reactions	Ethnicity	Reference
<i>Aromatic anticonvulsants</i>				
Carbamazepine	<i>B* 15:02</i>	SJS/TEN	Han Chinese, Thai, Indian, Malaysian, Vietnamese, Singaporean, Hong Kongese	[45, 82, 83, 226–230]
	<i>A* 31:01</i>	DRESS	Han Chinese, European, Spanish	[86, 87, 231]
	<i>A* 31:01</i>	DRESS/SJS/TEN	Northern European, Japanese, Korean	[88–90]
	<i>B* 15:11</i>	SJS/TEN	Han Chinese, Japanese, Korean	[89, 232, 233]
	<i>B* 59:01</i>	SJS/TEN	Japanese	[234]
	<i>B* 38:01</i>	SJS/TEN	Spanish	[231]
	<i>B* 15:02</i>	SJS/TEN	Han Chinese, Thai	[81, 84]
	<i>B* 15:02</i>	SJS/TEN	Han Chinese, Thai	[81, 83]
	<i>B* 15:02, B* 13:01, B* 51:01</i>	SJS/TEN	Han Chinese, Japanese, Malaysian	[91]
	<i>A* 33:03, B* 38:02, B* 51:01, B* 56:02, B* 58:01, C* 14:02</i>	SJS/TEN	Thai	[235]
Oxcarbazepine	<i>B* 51:01</i>	DRESS	Thai	[235]
	<i>B* 15:13</i>	DRESS/SJS/TEN	Malaysian	[236]
	<i>CYP2C9* 3</i>	DRESS/SJS/TEN	Han Chinese, Japanese, Malaysian	[91]
	<i>CYP2C9* 3</i>	SJS/TEN	Thai	[235]
	<i>B* 15:02</i>	SJS/TEN	Han Chinese	[81, 85, 237]
	<i>B* 38; B* 58:01, A* 68:01, Cw* 07:18</i>	SJS/TEN	European	[93, 238]
	Phenobarbital Lamotrigine	<i>B* 38:01</i>	SJS/TEN	Spanish
<i>A* 31:01</i>		SJS/TEN	Korean	[239]
<i>A* 24:02</i>		DRESS/SJS/TEN	Spanish	[231]
<i>Allopurinol</i>		<i>B* 58:01</i>	DRESS/SJS/TEN	Han Chinese, Thai, Japanese, Korean, European
	<i>Antiretroviral drugs</i>			
Abacavir	<i>B* 57:01</i>	HSS	European, African	[98, 99]
	<i>DRB1* 01:01</i>	DRESS	Australian	[240]
Nevirapine	<i>B* 35:05</i>	DRESS	Thai	[101]
	<i>B* 14:02, Cw* 08:01, Cw* 08:02</i>	HSS	Sardinian, Japanese	[102, 241]
	<i>C* 04:01</i>	DRESS/SJS/TEN	Malawian	[242]
<i>Antibiotics</i>				
Beta-lactam	<i>DR9, DR14.1, DR17, DR4</i>	Immediate-type drug hypersensitivity	Chinese	[243]
	<i>DRA rs7192, DRA rs8084</i>	Immediate-type drug hypersensitivity	Spanish, Italian	[66]
Cotrimoxazole	<i>B* 15:02, C* 06:02, C* 08:01</i>	SJS/TEN	Thai	[244]
Dapsone	<i>B* 13:01</i>	HSS	Han Chinese	[105]
Sulfamethoxazole	<i>B* 38:02</i>	SJS/TEN	European	[93]
Sulfonamide	<i>A* 29, B* 12, DR* 7</i>	TEN	European	[245]
<i>NSAIDs</i>				
Aspirin	<i>DRB1* 13:02, DRB1* 06:09</i>	Urticaria/angioedema	Korean	[74]
Aspirin and other NSAIDs	<i>DRB1* 11</i>	Urticaria/angioedema and hypotension/laryngeal edema	Spanish	[246]
Aspirin and other NSAIDs	<i>B* 44, Cw* 5</i>	Chronic idiopathic urticaria	Italian	[75]

TABLE 1: Continued.

Associated drug	HLA allele	Hypersensitivity reactions	Ethnicity	Reference
Oxicam NSAIDs	<i>B* 73:01</i>	SJS/TEN	European	[93]
<i>Other drugs</i>				
Methazolamide	<i>B* 59:01, CW* 01:02</i>	SJS/TEN	Korean, Japanese	[108]

DRESS: drug reaction with eosinophilia and systemic symptoms; HSS: hypersensitivity syndrome; MPE: maculopapular exanthema; NSAIDs: nonsteroidal anti-inflammatory drugs; SJS: Stevens-Johnson syndrome; TEN: toxic epidermal necrolysis.

TABLE 2: Genetic association with pathogenetic pathways in immediate-type drug hypersensitivity.

Associated drug	Ethnicity	Cytokines/chemokines	Production and release of mediators	Drug metabolism	Others	Reference
<i>Beta-lactam antibiotics</i>	Korean	—	<i>MS4A2</i>	—	—	[247, 248]
	Chinese	<i>IL4R, IL4, IL10, IL13, IFNGR1, STAT6</i>	—	—	—	[69, 70, 249–252]
	Italian	<i>IL4R, IL13, NOD2</i>	<i>LGALS3</i>	—	—	[66, 68, 73]
	French	<i>IL4R, IL10</i>	—	—	—	[253]
	American	<i>IL4R, IL4</i>	—	<i>LACTB</i>	—	[67]
	Spanish	<i>IL4R, TNF, NOD2</i>	<i>LGALS3</i>	—	—	[66, 73, 254, 255]
<i>Aspirin</i>	Korean	<i>IL18, TGFBI, TNF</i>	<i>ALOX5, FCER1A, FCER1G, HNMT, TBXA2R, PTGER4</i>	—	—	[76, 256–263]
	Poles	—	<i>LTC4S</i>	—	<i>GSTM1</i>	[264]
	Venezuelan	—	<i>LTC4S</i>	—	—	[265]
<i>NSAIDs</i>	Spanish	—	<i>ALOX5, ALOX5AP, ALOX15, CTSLTR1, DAO, PPARG, PTGDR, TBXAS1</i>	—	<i>CEP68</i>	[78, 266, 267]
	French	—	<i>ALOX5, PTGER1</i>	—	—	[268]
	Brazilian	<i>IL4R, IL10</i>	<i>DAO</i>	—	<i>CTLA4</i>	[269]

(CI) = 126–49,522) and has further been validated in cohorts of various other Asian populations including Thai, Indian, Malaysian, Vietnamese, Singaporean, and Hong Kongese cohorts [45, 80]. The *HLA-B*15:02* allele has also been identified as the common risk factor for SJS/TEN caused by other aromatic antiepileptic drugs [81], such as PHT [82, 83], OXC [84], and LTG [85]. The association between *HLA* alleles and CBZ-induced SCAR is phenotype and ethnic specific. The *HLA-A*31:01* allele is as specific predictor of CBZ-induced DRESS but not CBZ-induced SJS/TEN in Europeans and Han Chinese [86, 87]. In contrast, a strong association with *HLA-A*31:01* was found in CBZ-induced cutaneous adverse drug reactions (cADR) but not only in DRESS/DIHS in Northern Europeans, Japanese, and Koreans [88–90]. In addition to *HLA* alleles, a genome-wide association study showed a strong association of *CYP2C9*3* with PHT-induced SCAR in patients from Taiwan, Japan, and Malaysia and this finding was further supported by evidence indicating the delayed clearance of plasma PHT levels in PHT-induced SCAR [91].

3.2.2. Allopurinol. Allopurinol is a first-line drug used to treat gouty arthritis and urate nephropathy. In 2005, Hung et al. reported that *HLA-B*58:01* was the genetic risk marker for allopurinol-induced hypersensitivity in Han Chinese (corrected *P* value = 4.7×10^{-24} , OR = 580.3, and 95%

CI = 34.4–9780.9) [92]. This correlation was subsequently validated among different populations, including various Asian and European populations [93–96]. The gene dosage effect of *HLA-B*58:01* also influences the development of allopurinol-induced hypersensitivity (OR = 15.3 for *HLA-B*58:01* heterozygotes and OR = 72.5 for homozygotes), and the strength of the *HLA-B*58:01* association has been found to be correlated with the disease severity of allopurinol-induced hypersensitivity (OR = 8.5 for MPE, OR = 44.0 for SCAR) [97].

3.2.3. Antiretroviral Drugs, Antibiotics, and Other Drugs. The antiretroviral drugs, such as abacavir and nevirapine, are also known to cause hypersensitivity reactions. The association with abacavir was first found in 2002 due to the significant association between the *HLA-B*57:01* and abacavir-induced hypersensitivity reactions (corrected *P* value < 0.0001, OR = 117, and 95% CI = 29–481). The positive predictive value of *HLA-B*57:01* for abacavir hypersensitivity reactions has been reported to be 55% in Caucasians [98, 99]. Nevirapine, meanwhile, has been associated with nevirapine-induced hypersensitivity or DRESS in patients with *HLA-DRB1*01:01* in Western Australia [100], *HLA-B*35:05* in Thailand [101], and *HLA-Cw8* in Japan [102]. In addition, several antibiotic-induced hypersensitivity reactions and pharmacogenomic associations have also been reported,

such as sulfonamide-induced allergic reactions [103], penicillin-induced SCAR [104], *HLA-B*13:01* and dapsone-induced hypersensitivity syndrome in Chinese [105], *HLA-B*57:01* and flucloxacillin-induced liver injury [106], and *HLA-A*02:01* and *HLA-DQB1*06:02* and amoxicillin-clavulanate hepatitis [107]. Other pharmacogenomic associations include *HLA-B*59:01* and methazolamide-induced SJS/TEN in Koreans and Japanese [108], *HLA-B*73:01* and oxycam-induced SJS/TEN in Europeans [93], and *ABCB11*, *C-24T*, *UGT2B7*2*, and *IL-4 C-590-A* and diclofenac-induced liver disease in Europeans [109, 110].

4. Cellular Immunology and Immune Mechanisms in Drug Hypersensitivity

4.1. Antigen Presentation and Processing. Drugs are considered to be foreign antigens and bind to the HLA/peptide/TCR complex to trigger immune and hypersensitivity reactions. There are four hypotheses regarding drug presentation mechanisms that have been proposed to explain how small drug antigens might interact with HLA and TCR in drug hypersensitivity: (1) the hapten theory, (2) the pharmacological interaction with immune receptors (p-i) concept, (3) the altered peptide repertoire model, and (4) the altered TCR repertoire model [111–115].

First, the hapten theory states that the culprit drugs or their reactive metabolites are too small to be immunogenic on their own, whereas they covalently bind to the endogenous peptides to form an antigenic hapten-carrier complex. The hapten-carrier complex is presented to the HLA molecule and then recognized by TCR, resulting in the induction of drug-specific cellular or humoral immune responses. The hapten theory has been shown to be valid in cases of penicillin-induced cADR [111, 116]. Second, the pharmacological interaction with immune receptor (p-i) concept postulates that drugs may directly, reversibly, and noncovalently bind to the HLA and/or TCR protein and bypass the classic antigen-processing pathway in antigen-presenting cells. Wei et al. previously found that CBZ/aromatic antiepileptic drugs can directly interact with *HLA-B*15:02* protein. No intracellular antigen processing or drug metabolism was involved in the *HLA-B*15:02* presentation of CBZ [112]. Oxypurinol, the reactive metabolite of allopurinol, provides another example of the p-i concept in that it can directly and immediately activate drug-specific T cells via the preferential use of *HLA-B*58:01* without intracellular processing [113]. Third, the altered peptide repertoire model states that the culprit drugs occupy the position in the peptide-binding groove of the HLA protein, changing the binding cleft and the peptide specificity of HLA binding. Abacavir-induced hypersensitivity has been found to belong to this model, as the crystal structure of *HLA-B*57:01* has been found to form complexes with abacavir and peptides [114, 115]. These studies showed that abacavir binds to the F-pocket of *HLA-B*57:01* and alters the shape and chemistry of the antigen-binding cleft, thereby altering the repertoire of endogenous peptides and resulting in polyclonal T cell activation and autoimmune-like systemic reaction manifestations. Finally, the altered TCR repertoire model suggests

that some drugs, such as sulfamethoxazole, directly interact with TCR, but not with the peptides or HLA molecules. The drug antigens bind to specific TCRs and alter the conformation of those TCRs, giving them the potential to bind to HLA-self peptide complexes to elicit immune reactions [117]. In this model, TCR is regarded as an initial drug interaction molecule, suggesting that TCR is as crucial as HLA molecules and contributes to the occurrence of drug hypersensitivity. Furthermore, viruses have also been proposed to participate in HLA/drug/TCR interactions, in that they may provide exogenous peptides for drug presentation and play important roles in cADR [116].

4.2. Cellular Immunology and Immune Molecules Involved in Drug Hypersensitivity

4.2.1. Immediate-Type Drug Hypersensitivity. Immediate-type drug hypersensitivity can be mediated by IgE-mediated or non-IgE-mediated mechanisms [118]. IgE-mediated mechanisms are mediated by drug-specific IgE via an immune response to a hapten/carrier complex. In the primary drug sensitization, drug-specific IgE is formed when plasma cells are transformed from activated B cells and interact with T cells. In an allergic reaction, drug allergens bind to mast cells or basophils with high-affinity Fc receptors, to which drug-specific IgE is bound, causing degranulation of the mast cells or basophils that results in the release of various mediators, such as histamine, leukotrienes, prostaglandins, and cytokines [3]. Degranulation has recently been proposed to occur in two main forms that are related to reaction severity and progression: piecemeal degranulation and anaphylactic degranulation [2, 119]. Piecemeal degranulation is mediated through the upregulation of CD203c on basophils via the formation of small vesicles from the histamine-containing granules quickly shuttling to the plasma membrane to cause more severe and rapid reactions [120]. Anaphylactic degranulation results in the fusion of the main histamine-containing granules with the plasma membrane, releasing the entire contents of granules to the extracellular space and exposing CD63 on the surface of basophils [120].

The non-IgE-mediated immunologic mechanisms are mediated by IgG antibodies or by complement activation [23, 24]. IgG-mediated anaphylaxis has been established in mouse models, wherein the use of drugs with specific IgG bound to FcγRIII stimulates the release of platelet-activating factor (PAF) by basophils, macrophages, or neutrophils [24]. Although the IgG-mediated anaphylaxis mechanism has not been fully demonstrated in humans, some studies have shown that PAF is an essential mediator in such anaphylaxis [121]. In addition, a novel gain-of-function splice variant of FcγR FcγRIIA has been identified with the presence of IgG anti-IgA antibodies in patients with common variable immunodeficiency who developed anaphylaxis after intravenous immunoglobulin infusion [122]. Moreover, biological agents with IgA and infliximab have been shown to induce anaphylaxis in the absence of specific IgE but with high levels of specific IgG [123–125]. These observations also provide some additional evidence

for IgG-mediated anaphylaxis. Furthermore, complement activation can be induced through the absence of agent-specific IgE or IgG antibody immunocomplexes [24]. This condition can be observed in patients undergoing hemodialysis with a new dialysis membrane, protamine neutralization of heparin, and polyethylene glycol infusion [23, 126]. Drugs solubilized in therapeutic liposomes and lipid-based excipients (such as Cremophor EL used as the diluent for older preparations of propofol and paclitaxel) can form large micelles with serum lipids and cholesterol to stimulate the complement system [23, 126]. This activation of complement mechanisms further causes the release of C3a, C5a, and C5b-9, which trigger, in turn, the activation of mast cells, basophils, and other cells via their specific receptors, resulting in degranulation and mediator release [24].

The nonimmunologic-type hypersensitivity reaction directly activates mast cell degranulation without involving the activation of the immune system. There are several specific agents that induce different mechanisms beyond the direct immunoglobulin-mediated activation or complement activation. Oversulfated chondroitin sulfate-contaminated heparin was found to have caused various cases of anaphylaxis around 2007-2008 via the direct activation of the kinin system with increased production of bradykinin, C3a, and C5a [127]. The triggering of factor XII-driven contact system activation-mediated bradykinin formation also plays a key role in anaphylaxis [24]. NSAIDs, including aspirin, can result in anaphylactic reactions via the inhibition of cyclooxygenase with a decrease in the production of prostaglandins and the increased generation of cysteinyl leukotrienes [23]. Vancomycin can directly activate mast cells and/or basophils, leading to the release of histamine [128]. This mechanism was suggested to be mediated via the calcium-dependent activation of phospholipase-C and phospholipase-A2 pathways [128]. Opiates (e.g., meperidine, codeine, and morphine) also cause histamine release via direct mast cell degranulation [129]. Recently, it was proposed that nonimmunologic hypersensitivity reactions may also be mediated through the MAS-related G protein-coupled receptor-X2 (MRGPRX2) in cases involving specific drugs, such as icatibant, neuromuscular blocking drugs, and quinolone antibiotics [25]. The interaction of certain drugs with this mast cell receptor can stimulate degranulation and the release of TNF- α and prostaglandin D2 (PGD2), among other molecules, leading to nonimmunologic anaphylactic reactions [25]. The mouse counterpart of MRGPRX2 that participates in peptidergic drug-induced pseudoallergic reactions has been newly identified and could potentially be applied in preclinical screening models [25, 130].

4.2.2. Delayed-Type Drug Hypersensitivity. The main concept used to explain the pathomechanisms of delayed-type drug hypersensitivity consists of the view that specific T lymphocytes or natural killer (NK) cells are activated upon antigen recognition or Fas/FasL interaction and that various cytotoxic proteins, including perforin/granzyme B, and granulysin, are then released to attack keratinocytes or other cells, inducing skin rash or epidermal necrosis. In addition, several other cytokines/chemokines, including

TNF- α , IFN- γ , GM-CSF, TARC/CCL17, IL-6, IL-8/CXCL8, IL-15, and IL-36, are also known to participate in the immune reactions of drug hypersensitivity. These cytokines/chemokines have been found to be highly expressed in the skin lesions, blister fluids, blister cells, peripheral blood mononuclear cells (PBMC), or plasma of patients. These immune mediators are responsible for the trafficking, proliferation, regulation, or activation of T lymphocytes and other leukocytes, thereby affecting the clinical presentations of drug hypersensitivity in various ways (Table 3).

(1) *Fas-FasL Interaction.* Fas ligand (FasL) belongs to the tumor necrosis factor (TNF) family. The binding of Fas and FasL plays an important role in regulating the immune system and is involved in the apoptosis of epidermal cells in patients with drug hypersensitivity. Briefly, upon Fas-FasL interaction, the Fas-associated death domain protein (FADD) is recruited and binds to the Fas-FasL complex. The FADD then recruits procaspase 8, bringing multiple copies of procaspase 8 together, which in turn autoactivate to become caspase 8, triggering the caspase cascade and resulting in intracellular DNA degradation [131]. Viard et al. proposed that a suicidal interaction between Fas and FasL, which are both expressed by keratinocytes, leads to the extensive necrosis of epidermal cells in individuals with SJS/TEN [132].

(2) *Perforin/Granzyme B.* A controversial hypothesis suggests that perforin and granzyme B play more important roles in the keratinocyte death in SJS/TEN than does the Fas-FasL interaction [133]. Granzymes are serine proteases that are released by cytoplasmic granules and can induce programmed cell death in the target cells. Upon activation, drug-specific cytotoxic T lymphocytes (CTL) and NK cells produce perforin, which can bind to and punch a channel through the cell membrane, promoting the entry of granzyme B into the target cells to activate the caspase cascade and the succeeding apoptosis [134]. Delayed reactions to drugs have shown that increasing levels of perforin and granzyme B are related to the disease severity of drug hypersensitivity [131].

(3) *Granulysin.* Granulysin is a cytolytic protein mainly released by CTL and NK cells. It functions to create holes in the cell membranes and thereby destroy target cells. In 2008, Chung et al. reported that 15 kDa secretory granulysin serves as a key mediator for the disseminated keratinocyte apoptosis seen in SJS/TEN [135]. In that study, the increased level of granulysin in blister fluids from the skin lesions of SJS/TEN patients was much higher than the levels of other cytotoxic proteins, such as perforin, granzyme B, and FasL, and depleting the granulysin reduced the cytotoxicity [135]. Further studies demonstrated that granulysin is strongly expressed in patients with drug-induced FDE, DRESS/DIHS, and SJS/TEN but not MPE [136-138].

(4) *TNF- α , IFN- γ , TARC, IL-15, and Other Cytokines/Chemokines in SJS/TEN, DRESS/DIHS, and AGEP.* TNF- α is a major proinflammatory cytokine and is produced by

TABLE 3: Delayed-type drug hypersensitivity-related cytokines and chemokines.

Phenotype	Cytokines/chemokines	Skin or blister	Plasma	PBMC	References
DRESS/DIHS	TNF- α		+		[160]
	IFN- γ	+	+	+	[270–272]
	IL-2			+	[270]
	IL-4			+	[270]
	IL-5			+	[270]
	IL-6		+		[160]
	IL-13			+	[270]
	IL-15			+	[138]
	TARC/CCL17		+		[273]
SJS/TEN	TNF- α	+	+	+	[131, 138, 141–143, 274, 275]
	IFN- γ	+		+	[131, 142, 143, 274]
	IL-2	+		+	[131, 143]
	IL-5	+			[143]
	IL-6	+	+	+	[143, 153, 154, 138]
	IL-8/CXCL8		+		[138]
	IL-10	+	+	+	[142, 153]
	IL-12	NS			[142]
	IL-13	+			[143]
	IL-15	NS		+	[142, 138]
	IL-18	+			[142]
	CCR3	+			[143]
CXCR3	+			[143]	
CXCR4	NS			[143]	
	CCR10			+	[152]
AGEP	IL-8/CXCL8	+			[145, 146]
	IL-36	+			[147, 148]
	GM-CSF			+	[145]

AGEP: acute generalized exanthematous pustulosis; CCR: C–C chemokine receptor; CXCR: CX chemokine receptor; DIHS: drug-induced hypersensitivity syndrome; DRESS: drug reactions with eosinophilia and systemic symptoms; IFN- γ : interferon- γ ; IL: interleukin; NS: not significant; SJS/TEN: Stevens–Johnson syndrome and toxic epidermal necrolysis; TNF- α : tumor necrosis factor- α .

macrophages, T lymphocytes, NK cells, neutrophils, mast cells, and eosinophils. It regulates immune responses through the induction of cell apoptosis, activation, differentiation, and inflammation [139]. TNF- α was highly expressed and suggested to be responsible for the extensive necrosis of skin lesions of SCAR patients [140, 141]. IFN- γ is critical for both innate and adaptive immunity against viral and bacterial infection, and it is predominantly produced by CD4⁺ T helper cells, CD8⁺ CTL, and NK cells. IFN- γ was found to be increased in the skin tissue, blister cells, and plasma of patients with erythema multiforme, SJS, TEN, and DRESS/DIHS [131, 142, 143]. The immune mechanism of AGEP is not yet well understood. However, high levels of IL-8/CXCL8 production and the recruitment of neutrophils have been observed in the skin lesions of AGEP patients [144–146]. Mutations in the *IL36RN* gene encoding the IL-36 receptor antagonist (IL-36Ra) have also been identified in AGEP patients [147, 148]. DRESS/DIHS is characterized by leukocytosis with atypical lymphocytosis or eosinophilia [149]. Serum thymus and activation-regulated chemokine (TARC) was identified as a potential biomarker for early

indication of the disease and a predictor of disease activity in DRESS/DIHS [150, 151]. Compared to patients with MPE and SJS/TEN, the TARC levels in patients with DRESS/DIHS are significantly higher during the acute phase and are correlated with skin eruptions [151]. Interleukin-15 (IL-15) is a cytokine that can induce the proliferation of NK cells and other leukocytes, and it has been found to be associated with the disease severity and mortality of SJS/TEN [138]. IL-15 has also been shown to enhance the cytotoxicity of cultured NK cells and blister cells from TEN patients [138]. In addition, other cytokines and chemokine receptors, including IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-18, CCR3, CXCR3, CXCR4, and CCR10, have been found to be upregulated in the skin lesions, blister fluids, PBMC, or plasma of drug hypersensitivity patients and to participate in the immune regulation of drug hypersensitivity [131, 138, 142, 143, 152–154].

(5) *Syndrome-Specific Effector Cells*. SJS/TEN is characterized by profound necrosis localized to the epidermis. Cytotoxic CD8 T cells, natural killer cells, and natural killer T cells

producing the cytotoxic molecules, especially granulysin, which causes extensive keratinocyte death, are enriched in blister fluid samples from the skin lesions of patients with SJS/TEN. Granulysin serum levels are correlated with the severity of acute disease and mortality [135, 155]. These cytotoxic cells mediate the disease pathogenesis. It is shown that the function of regulatory T cells (Tregs) in SJS/TEN is inadequate, although present in normal frequency [156]. Immunological changes of DRESS/DIHS are characterized by the increase of atypical lymphocytes or eosinophils [149, 157]. Eosinophilia can be observed in 60–95% of DRESS/DIHS patients at the early stage of the illness [32, 157]. Most of DRESS patients had increased numbers of CD4⁺ T cells in the acute stage, which was associated with the severity of clinical symptoms, such as the extent of skin rash and reactivations of virus [158]. In addition, Tregs play important roles in DRESS/DIHS pathogenesis. Dramatic expansions of functional Tregs are found in the acute stage of DRESS/DIHS [156]. It is hypothesized that CD4⁺FoxP3⁺ T cells that are home to skin serve to limit the severity of acute disease by regulating the cytotoxic effector T cell responses. However, Treg responses eventually exhaust and this might contribute to ongoing viral replication and intermittent recurrence of clinical symptoms [156, 159]. In patients with AGEP, it is shown that the increased neutrophilic inflammatory processes are regulated by T lymphocytes, which is important in the pathogenesis. The recruitment of neutrophils was observed in the skin lesions of the patients with the late phase of disease development [144, 145].

5. Environmental Factors and Viral Infections in Drug Hypersensitivity

In addition to drug antigens, hypersensitivity reactions may be induced by other pathogens, such as *Mycoplasma pneumoniae*, or viral infections. Virus-drug interactions associated with viral reactivation may also exist. For example, it is well known that human herpesvirus-6 (HHV-6) plays an important role in DRESS/DIHS. HHV-6 reactivation in patients with DRESS/DIHS may increase T cell activity after the initiation of the drug eruption and induce the synthesis of proinflammatory cytokines, including TNF- α and IL-6, which may in turn modulate the T cell-mediated responses [160]. Shiohara et al. reviewed the associations between viral infections and drug rashes, as well as the mechanisms by which viral infections induce drug rashes. The sequential reactivations of several herpes viruses (HHV-6, HHV-7, Epstein-Barr virus (EBV), and cytomegalovirus (CMV)) were found to be coincident with the clinical symptoms of drug hypersensitivity reactions [161]. Chung et al. reported that a new variant of coxsackievirus A6 (CVA6) acting as the causative agent may induce widespread mucocutaneous blistering reactions mimicking the features of erythema multiforme major or SCAR [52]. In addition, the virus may also provide exogenous peptides for drug presentation and participate in HLA/drug/TCR interactions. White et al. recently proposed that some patients may acquire primary infection via HHVs or other pathogens that in turn induce drug hypersensitivity [116]. The presence of HHV peptides in

patients with high-risk HLA alleles may trigger the activation of cytotoxic T cells, thereby resulting in the development of SCAR. The pathogenic factors underlying the unusual presentations of drug hypersensitivity related to viral infections need to be further investigated.

6. Diagnostic Tools for Drug Hypersensitivity

6.1. Diagnostic Tools for Immediate-Type Drug Hypersensitivity. The most commonly used laboratory test for confirming a diagnosis of anaphylaxis consists of determining the patient's total serum tryptase level [162]. Serial measurements of tryptase levels can be taken during an anaphylactic episode, although measurements of the baseline level are considered to be most useful. In fact, while serial measurements of tryptase levels taken during an anaphylactic episode can serve as useful markers for evaluating these reactions, this approach is not used so widely in clinical practice due to the limitations involved in measuring tryptase during the acute phase of an episode. Elevated levels of histamine, the first mediator released by mast cells, in plasma or urine are also consistent with anaphylaxis [2]. However, plasma histamine levels are only transiently elevated, making them of little utility if the patient is evaluated more than 1 hour after onset of the episode [163]. At the same time, normal levels of tryptase or histamine do not preclude a diagnosis of drug hypersensitivity [15]. Other newly identified biomarkers, such as PAF and carboxypeptidase A3, bring hope for enhancing diagnostic accuracy, although their use remains experimental [15, 164].

For IgE-mediated hypersensitivity reactions, serum drug-specific IgE (sIgE) quantification and the basophil activation test (BAT) are frequently used to assess the culprit drug. The tests used to conduct sIgE immunoassays consist of radioallergen sorbent testing (RAST), enzyme-linked immunosorbent assays (ELISAs), and fluoroenzyme immunoassays (FEIAs) [165]. While RAST or ELISAs are usually conducted using in-house techniques, FEIAs can be performed using commercial products, such as the ImmunoCAP-FEIA system [166–168]. Only a few products are available, meanwhile, for some drugs, particularly beta-lactam antibiotics [167, 169]. The sensitivity of the various immunoassays used has been found to average 62.9%, while the average specificity, PPV, and NPV are 89.2%, 83.3%, and 77.8%, respectively [168]. The average NPV is also relatively low in order to exclude allergic reactions and determine whether to perform a provocation test [170]. In comparison, the BAT test provides a higher average specificity (94.6%) and PPV (93.4%) than immunoassays [168]. The test uses flow cytometry after drug stimulation to determine the levels of basophil activation or degranulation markers; the upregulation of CD63 and CD203c is also usually measured [171]. Of note, the results of the BAT for aspirin/NSAID-induced hypersensitivity remain inconclusive due to the fact that they encompass both IgE-mediated allergic reactions and nonimmunological intolerances, limiting the use of the BAT in assessing non-IgE-mediated reactions [172]. Mediator release assays, meanwhile, measure the mediator released (histamine or leukotriene 4) in a supernatant upon cell activation after

drug stimulation, but these assays have exhibited sensitivity and specificity levels too low for them to be recommended for the purposes of diagnosis [169, 173].

6.2. Diagnostic Tools for Delayed-Type Drug Hypersensitivity. The discovery of biomarkers for drug hypersensitivity is crucial for clinical purposes, including the early diagnosis and better prediction of this disease in order to prevent complications. We previously found granulysin to be a key cytotoxic molecule responsible for disseminated keratinocyte necrosis through the action of cytotoxic lymphocytes or NK-cell-mediated cytotoxicity with no direct cellular contact [135]. A significant correlation between the granulysin levels in blister fluids and clinical severity was also found [135]. In addition, the serum granulysin levels in patients with SJS/TEN have also been found to be significantly elevated before the development of skin detachment or mucosal lesions but then to drop rapidly within 5 days of disease onset [136]. As a potential marker for the early phase of SJS/TEN, a simple rapid immunochromatographic test for elevated serum granulysin was developed for immediate clinical use. Additionally, prolonged elevation of serum granulysin has also been found in DIHS patients, indicating that such elevation could possibly be used for the purposes of early diagnosis and predicting disease prognosis [174]. Furthermore, the levels of IL-15 were correlated with the disease progression and mortality of SJS/TEN at early stage [138]. Serum IL-15 levels can be further utilized as a marker for early diagnosis and prognosis monitoring [138]. For DRESS/DIHS, serum TARC levels in patients with DRESS/DIHS have been reported to be significantly higher than those in patients with SJS/TEN and MPE during the acute phase and to be correlated with skin eruptions [151]. TARC was thus identified as a potential biomarker for the early indication and disease activity of DRESS/DIHS and also for determining the prognosis of systemic severity of inflammation in drug eruptions other than SJS/TEN [150, 151]. For AGEF, meanwhile, no specific markers for diagnosing or predicting the disease have been identified at present [175].

Drug rechallenge is considered the gold standard for confirming a potential offending drug; however, its use is not practical due to the possible life-threatening consequences. As such, there is still no standard method for the confirmation of drug causality. Nonetheless, since HLA genotyping has been useful in screening for populations at risk for SCAR, HLA genotyping might be helpful for identifying culprit drugs via specific HLA alleles in at-risk populations [48, 176]. Several *in vitro* tests can be used to assist in the confirmation of drug causality, but the exact sensitivity and specificity of such tests are not well known [177, 178]. There are several tests currently available: the lymphocyte transformation test (LTT), ELISpot (Enzyme-linked immunospot assay) intracellular cytokine staining, and the enzyme-linked immunosorbent assay (ELISA) for the secretion of cytotoxic mediators including inflammatory cytokines, chemokine-chemokine receptors, IFN- γ , Fas-Fas ligand, perforin, granzyme B, and granulysin [179]. The LTT is a reproducible test for measuring the enhanced proliferative response of PBMC after the sensitization of T cells to a drug

[180]. However, the sensitivity of the test has reportedly varied among various studies involving various drugs and clinical phenotypes and different timings for use of the test [181, 182]. The relevance of using the LTT in testing for SJS/TEN was relatively lower than those using than DRESS/DIHS and AGEF [182]. Several modifications can help to increase the sensitivity of the LTT or ELISpot, including stimulation with anti-CD-3/CD28 antibody-coated microbeads with IL-2, depletion of Treg/CD25hi cells, or the combined addition of anti-CTLA4 and anti-programmed cell death ligand 1 (PD-L1) antibodies to PBMC cultures [183–185]. IFN- γ -ELISpot showed a similar sensitivity (67%) and specificity in DRESS, but a higher sensitivity (71%) in SJS/TEN [179]. The data for an ELISA-based test used to detect granulysin showed better sensitivity (86%) in SJS/TEN, but the evidence was limited due to the small number of cases in the study [186]. Further larger studies will thus be needed to confirm both the sensitivity and specificity.

In vivo patch tests provide a low-risk method for reproducing delayed hypersensitivity with moderate reexposure of patients to suspected offending drugs [187]. The value of patch testing depends on the phenotypes and drugs involved. The sensitivity of such testing is generally <70%, but higher sensitivities have been reported for AGEF and for some selected populations such as abacavir-hypersensitivity, carbamazepine-induced SJS/DRESS, and fixed drug eruption patients [178, 187, 188]. The skin tests involving a prick or intradermal testing are considered to be crucial tools for evaluating drug hypersensitivity reactions, including IgE-mediated or delayed-type hypersensitivity, in both the European and American guidelines [22, 189–191]. However, these skin tests are usually not suggested for SCAR patients due to the risk of relapse, although late-reading intradermal tests are of value for AGEF patients and negative patch tests are of value for SCAR patients [187, 192].

7. Therapeutic Approaches in Drug Hypersensitivity

7.1. Therapeutic Approaches in Immediate-Type Drug Hypersensitivity. Anaphylaxis is a medical emergency and epinephrine is the treatment of choice for anaphylaxis to prevent its progression to a life-threatening condition [15, 193]. Epinephrine should be administered as soon as possible without delay to avoid mortality [194]. The intramuscular injection of epinephrine into the middle of the outer thigh is recommended to treat anaphylaxis in most settings and in patients of all ages [195]. Glucagon is indicated for patients receiving beta-blockers with refractory symptoms [196]. The use of corticosteroids was previously believed to decrease the risk of biphasic and protracted reactions; however, a systematic review of the literature failed to retrieve any randomized controlled trials to confirm their effectiveness [197]. An emergency department-based study also failed to find a decrease in the rates of return visits or biphasic reactions among patients treated with glucocorticoids [198]. These adjunctive therapies, including corticosteroids, antihistamines, and bronchodilators, could help to relieve symptoms,

but should not be substituted for epinephrine or delay the use of epinephrine [199, 200].

7.2. Therapeutic Approaches in Delayed-Type Drug Hypersensitivity. For the treatment of severe delayed-type drug hypersensitivity, such as SJS/TEN, there are no optimal treatment guidelines. Thus far, in fact, only a few randomized trials that could be regarded as references to guide treatment have been conducted. The efficacy of systemic immunosuppressants or immunomodulatory treatments (e.g., corticosteroids, cyclosporine, intravenous immunoglobulins (IVIg), and plasmapheresis) still remains controversial. Systemic corticosteroids could be the most common treatment option, but the prior use of corticosteroids was found to prolong disease progression with no definite benefit in terms of survival [60, 201–203]. IVIg is one of the most commonly utilized therapies for SJS/TEN and is frequently the adjunctive therapy used for severe cases or pediatric patients [204]. In a meta-analysis, however, IVIg, even high doses of IVIg, failed to achieve statistically significant results supporting the conclusion that it is clinically beneficial [204, 205]. IVIg has been found to yield better outcomes in pediatric patients, but children with TEN usually have lower rates of mortality and better prognoses than adult patients [204, 206]. Cyclosporine, has been found to decrease the mortality rate and the progression of detachment in adults in an open-label phase II trial [207]. However, one recent cohort study revealed a statistically insignificant survival benefit for cyclosporine therapy compared to supportive care [208]. In contrast, the first meta-analysis of 7 studies regarding the effect on mortality of cyclosporine in the treatment of SJS/TEN showed a beneficial effect [209]. A trend identified in the same study also indicated that cyclosporine demonstrated better survival than IVIg [209]. There have also been an increasing number of case reports regarding the benefit of treatment with anti-TNF- α biologic agents for patients with TEN [210–215]. One recent systemic review showed that glucocorticosteroids and cyclosporine are the most promising therapies in terms of survival benefit, but no such benefits were observed for IVIg, plasmapheresis, thalidomide, cyclophosphamide, hemoperfusion, tumor necrosis factor inhibitors, or granulocyte colony-stimulating factor [216]. Meanwhile, IL-15 was demonstrated to be a major cytokine orchestrating SJS/TEN, indicating that further novel therapeutics including IL-15 blockers, the mammalian target of rapamycin (mTOR) inhibitors, and Janus kinase/signal transducers and activators of transcription (JAK/STAT) inhibitors hold promise for impacting various therapeutic targets [138, 217]. That said, further prospective, randomized controlled studies are needed to provide more definitive conclusions regarding treatment in patients with SJS/TEN.

Systemic corticosteroids have been considered the treatment of choice for patients with DRESS/DIHS, but they may be associated with an increased risk of complications such as opportunistic infections [218]. CMV and HHV-6 viral loads were also reported to be increased in patients receiving systemic corticosteroids, while EBV loads were higher in patients not receiving systemic corticosteroids

[219]. Antiviral medications such as ganciclovir can be given in addition to steroids and/or IVIg in cases of severe disease with confirmation of viral reactivation [220]. Several previous studies have reported the effectiveness of treatment with IVIg [221]. However, the premature discontinuation of a prospective study regarding the role of IVIg treatment occurred due to severe adverse effects [222]. Plasmapheresis and other immunosuppressive drugs, such as cyclophosphamide, cyclosporine, interferons, muromonab-CD3, mycophenolate mofetil, and rituximab, may also be potential therapies [221]. Among the above treatments, the use of cyclosporine was successful in 2 recent cases with rapid response, and so, its use could be considered for patients with concerns about using longer courses of systemic corticosteroids [223]. Supportive treatment with topical steroid-based treatments for AGEF is suggested due to the mostly benign and self-limiting course of the condition [224, 225]. Meanwhile, the administration of systemic steroids for a short period can be considered for severe and refractory cases [175].

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this paper.

Authors' Contributions

Yi-Giien Tsai and Wen-Hung Chung contributed equally to this work.

Acknowledgments

The authors would like to thank the National Science Council, Taiwan (MOST103-2321-B-182-001, MOST101-2628-B-182-001-MY3, MOST104-2314-B-182A-148-MY3, and MOST104-2325-B-182A-006), and Chang Gung Memorial Hospital (CLRPG2E0051~3, CORPG3F0041~2, OMRPG3E0041, and CMRPG1F0111~2) for kindly supporting this work.

References

- [1] W. J. Pichler, "Delayed drug hypersensitivity reactions," *Annals of Internal Medicine*, vol. 139, no. 8, pp. 683–693, 2003.
- [2] M. I. Montañez, C. Mayorga, G. Bogas et al., "Epidemiology, mechanisms, and diagnosis of drug-induced anaphylaxis," *Frontiers in Immunology*, vol. 8, p. 614, 2017.
- [3] B. Schnyder and W. J. Pichler, "Mechanisms of drug-induced allergy," *Mayo Clinic Proceedings*, vol. 84, no. 3, pp. 268–272, 2009.
- [4] S. G. Johansson, T. Bieber, R. Dahl et al., "Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003," *The Journal of Allergy and Clinical Immunology*, vol. 113, no. 5, pp. 832–836, 2004.
- [5] M. Mockenhaupt, "Epidemiology of cutaneous adverse drug reactions," *Chemical Immunology and Allergy*, vol. 97, pp. 1–17, 2012.

- [6] G. A. Wong and N. H. Shear, "Adverse drug interactions and reactions in dermatology: current issues of clinical relevance," *Dermatologic Clinics*, vol. 23, no. 2, pp. 335–342, 2005.
- [7] C. C. Vittorio and J. J. Muglia, "Anticonvulsant hypersensitivity syndrome," *Archives of Internal Medicine*, vol. 155, no. 21, pp. 2285–2290, 1995.
- [8] F. Arellano and J. A. Sacristan, "Allopurinol hypersensitivity syndrome: a review," *Annals of Pharmacotherapy*, vol. 27, no. 3, pp. 337–343, 1993.
- [9] H. Bocquet, M. Bagot, and J. C. Roujeau, "Drug-induced pseudolymphoma and drug hypersensitivity syndrome (drug rash with eosinophilia and systemic symptoms: DRESS)," *Seminars in Cutaneous Medicine and Surgery*, vol. 15, no. 4, pp. 250–257, 1996.
- [10] T. Shiohara, M. Iijima, Z. Ikezawa, and K. Hashimoto, "The diagnosis of a DRESS syndrome has been sufficiently established on the basis of typical clinical features and viral reactivations," *British Journal of Dermatology*, vol. 156, no. 5, pp. 1083–1084, 2007.
- [11] Y. Ushigome, Y. Kano, K. Hirahara, and T. Shiohara, "Human herpesvirus 6 reactivation in drug-induced hypersensitivity syndrome and DRESS validation score," *The American Journal of Medicine*, vol. 125, no. 7, pp. e9–e10, 2012.
- [12] S. Bastuji-Garin, B. Rzany, R. S. Stern, N. H. Shear, L. Naldi, and J. C. Roujeau, "Clinical classification of cases of toxic epidermal necrolysis, Stevens-Johnson syndrome, and erythema multiforme," *Archives of Dermatology*, vol. 129, no. 1, pp. 92–96, 1993.
- [13] A. Sidoroff, A. Dunant, C. Viboud et al., "Risk factors for acute generalized exanthematous pustulosis (AGEP)-results of a multinational case-control study (EuroSCAR)," *British Journal of Dermatology*, vol. 157, no. 5, pp. 989–996, 2007.
- [14] M. Lerch and W. J. Pichler, "The immunological and clinical spectrum of delayed drug-induced exanthems," *Current Opinion Allergy and Clinical Immunology*, vol. 4, no. 5, pp. 411–419, 2004.
- [15] F. E. Simons, L. R. Arduoso, M. B. Bilò et al., "World Allergy Organization anaphylaxis guidelines: summary," *The Journal of Allergy and Clinical Immunology*, vol. 127, no. 3, pp. 587–593.e22, 2011.
- [16] E. J. Jares, C. E. Baena-Cagnani, M. Sánchez-Borges et al., "Drug-induced anaphylaxis in Latin American countries," *The Journal of Allergy and Clinical Immunology: in Practice*, vol. 3, no. 5, pp. 780–788, 2015.
- [17] Y. S. Lee and W. Z. Sun, "Epidemiology of anaphylaxis: a retrospective cohort study in Taiwan," *Asian Journal of Anesthesiology*, vol. 55, no. 1, pp. 9–12, 2017.
- [18] L. H. Garvey, "Old, new and hidden causes of perioperative hypersensitivity," *Current Pharmaceutical Design*, vol. 22, no. 45, pp. 6814–6824, 2016.
- [19] M. Iammatteo, T. Keskin, and E. Jerschow, "Evaluation of periprocedural hypersensitivity reactions," *Annals of Allergy Asthma & Immunology*, vol. 119, no. 4, pp. 349.e2–355.e2, 2017.
- [20] N. Blanca-Lopez, D. Perez-Alzate, I. Andreu et al., "Immediate hypersensitivity reactions to ibuprofen and other arylpropionic acid derivatives," *Allergy*, vol. 71, no. 7, pp. 1048–1056, 2016.
- [21] M. G. Canto, I. Andreu, J. Fernandez, and M. Blanca, "Selective immediate hypersensitivity reactions to NSAIDs," *Current Opinion in Allergy and Clinical Immunology*, vol. 9, no. 4, pp. 293–297, 2009.
- [22] K. Brockow, L. H. Garvey, W. Aberer et al., "Skin test concentrations for systemically administered drugs - an ENDA/EAACI Drug Allergy Interest Group position paper," *Allergy*, vol. 68, no. 6, pp. 702–712, 2013.
- [23] R. Munoz-Cano, C. Picado, A. Valero, and J. Bartra, "Mechanisms of anaphylaxis beyond IgE," *Journal of Investigational Allergology and Clinical Immunology*, vol. 26, no. 2, pp. 73–82, 2016.
- [24] F. D. Finkelman, M. V. Khodoun, and R. Strait, "Human IgE-independent systemic anaphylaxis," *The Journal of Allergy and Clinical Immunology*, vol. 137, no. 6, pp. 1674–1680, 2016.
- [25] H. Subramanian, K. Gupta, and H. Ali, "Roles of Mas-related G protein-coupled receptor X2 on mast cell-mediated host defense, pseudoallergic drug reactions, and chronic inflammatory diseases," *The Journal of Allergy and Clinical Immunology*, vol. 138, no. 3, pp. 700–710, 2016.
- [26] D. Laroche, F. Namour, C. Lefrancois et al., "Anaphylactoid and anaphylactic reactions to iodinated contrast material," *Allergy*, vol. 54, Supplement 58, pp. 13–16, 1999.
- [27] K. Farnam, C. Chang, S. Teuber, and M. E. Gershwin, "Nonallergic drug hypersensitivity reactions," *International Archives of Allergy and Immunology*, vol. 159, no. 4, pp. 327–345, 2012.
- [28] M. L. Kowalski, R. Asero, S. Bavbek et al., "Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs," *Allergy*, vol. 68, no. 10, pp. 1219–1232, 2013.
- [29] S. H. Kardaun, A. Sidoroff, L. Valeyrie-Allanore et al., "Variability in the clinical pattern of cutaneous side-effects of drugs with systemic symptoms: does a DRESS syndrome really exist?," *British Journal of Dermatology*, vol. 156, no. 3, pp. 609–611, 2007.
- [30] J. R. Sullivan and N. H. Shear, "The drug hypersensitivity syndrome: what is the pathogenesis?," *Archives of Dermatology*, vol. 137, no. 3, pp. 357–364, 2001.
- [31] P. Cacoub, P. Musette, V. Descamps et al., "The DRESS syndrome: a literature review," *The American Journal of Medicine*, vol. 124, no. 7, pp. 588–597, 2011.
- [32] S. H. Kardaun, P. Sekula, L. Valeyrie-Allanore et al., "Drug reaction with eosinophilia and systemic symptoms (DRESS): an original multisystem adverse drug reaction. Results from the prospective RegiSCAR study," *British Journal of Dermatology*, vol. 169, no. 5, pp. 1071–1080, 2013.
- [33] C. H. Wei, R. Chung-Yee Hui, C. J. Chang et al., "Identifying prognostic factors for drug rash with eosinophilia and systemic symptoms (DRESS)," *European Journal of Dermatology*, vol. 21, no. 6, pp. 930–937, 2011.
- [34] J. C. Roujeau and R. S. Stern, "Severe adverse cutaneous reactions to drugs," *The New England Journal of Medicine*, vol. 331, no. 19, pp. 1272–1285, 1994.
- [35] Y. Kano, T. Ishida, K. Hirahara, and T. Shiohara, "Visceral involvements and long-term sequelae in drug-induced hypersensitivity syndrome," *Medical Clinics of North America*, vol. 94, no. 4, pp. 743–759, 2010.
- [36] O. Matsuno, "Drug-induced interstitial lung disease: mechanisms and best diagnostic approaches," *Respiratory Research*, vol. 13, no. 1, p. 39, 2012.

- [37] T. Thongsri, L. Chularojanamontri, and W. J. Pichler, "Cardiac involvement in DRESS syndrome," *Asian Pacific Journal Allergy and Immunology*, vol. 35, no. 1, pp. 3–10, 2017.
- [38] G. P. Bourgeois, J. A. Cafardi, V. Groysman, and L. C. Hughey, "A review of DRESS-associated myocarditis," *Journal of the American Academy of Dermatology*, vol. 66, no. 6, pp. e229–e236, 2012.
- [39] M. Ben m'rad, S. Leclerc-Mercier, P. Blanche et al., "Drug-induced hypersensitivity syndrome: clinical and biologic disease patterns in 24 patients," *Medicine*, vol. 88, pp. 131–140, 2009.
- [40] J. C. Roujeau, J. C. Guillaume, J. P. Fabre, D. Penso, M. L. Flechet, and J. P. Girre, "Toxic epidermal necrolysis (Lyell syndrome). Incidence and drug etiology in France, 1981–1985," *Archives of Dermatology*, vol. 126, no. 1, pp. 37–42, 1990.
- [41] E. Schopf, A. Stuhmer, B. Rzany, N. Victor, R. Zentgraf, and J. F. Kapp, "Toxic epidermal necrolysis and Stevens-Johnson syndrome. An epidemiologic study from West Germany," *Archives of Dermatology*, vol. 127, no. 6, pp. 839–842, 1991.
- [42] D. Y. Hsu, J. Brieva, N. B. Silverberg, and J. I. Silverberg, "Morbidity and mortality of Stevens-Johnson syndrome and toxic epidermal necrolysis in United States adults," *Journal of Investigative Dermatology*, vol. 136, no. 7, pp. 1387–1397, 2016.
- [43] N. Frey, J. Jossi, M. Bodmer et al., "The epidemiology of Stevens-Johnson syndrome and toxic epidermal necrolysis in the UK," *Journal of Investigative Dermatology*, vol. 137, no. 6, pp. 1240–1247, 2017.
- [44] J. Diphoorn, S. Cazzaniga, C. Gamba et al., "Incidence, causative factors and mortality rates of Stevens-Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN) in northern Italy: data from the REACT registry," *Pharmacoeconomics and Drug Safety*, vol. 25, no. 2, pp. 196–203, 2016.
- [45] W. H. Chung, S. I. Hung, H. S. Hong et al., "Medical genetics: a marker for Stevens-Johnson syndrome," *Nature*, vol. 428, no. 6982, p. 486, 2004.
- [46] D. Y. Hsu, J. Brieva, N. B. Silverberg, A. S. Paller, and J. I. Silverberg, "Pediatric Stevens-Johnson syndrome and toxic epidermal necrolysis in the United States," *Journal of the American Academy of Dermatology*, vol. 76, no. 5, pp. 811.e4–817.e4, 2017.
- [47] P. B. Ferrell Jr. and H. L. McLeod, "Carbamazepine, HLA-B*1502 and risk of Stevens-Johnson syndrome and toxic epidermal necrolysis: US FDA recommendations," *Pharmacogenomics*, vol. 9, no. 10, pp. 1543–1546, 2008.
- [48] E. J. Phillips, W. H. Chung, M. Mockenhaupt, J. C. Roujeau, and S. A. Mallal, "Drug hypersensitivity: pharmacogenetics and clinical syndromes," *The Journal of Allergy and Clinical Immunology*, vol. 127, no. 3, pp. S60–S66, 2011.
- [49] A. Auquier-Dunant, M. Mockenhaupt, L. Naldi et al., "Correlations between clinical patterns and causes of erythema multiforme majus, Stevens-Johnson syndrome, and toxic epidermal necrolysis - results of an international prospective study," *Archives of Dermatology*, vol. 138, no. 8, pp. 1019–1024, 2002.
- [50] G. Yildirim Cetin, H. Sayar, F. Ozkan, S. Kurtulus, F. Kesici, and M. Sayarlioglu, "A case of toxic epidermal necrolysis-like skin lesions with systemic lupus erythematosus and review of the literature," *Lupus*, vol. 22, no. 8, pp. 839–846, 2013.
- [51] D. Olson, L. K. Watkins, A. Demirjian et al., "Outbreak of mycoplasma pneumoniae-associated Stevens-Johnson syndrome," *Pediatrics*, vol. 136, no. 2, pp. e386–e394, 2015.
- [52] W. H. Chung, S. R. Shih, C. F. Chang et al., "Clinicopathologic analysis of coxsackievirus a6 new variant induced widespread mucocutaneous bullous reactions mimicking severe cutaneous adverse reactions," *The Journal of Infectious Diseases*, vol. 208, no. 12, pp. 1968–1978, 2013.
- [53] M. Mockenhaupt, C. Viboud, A. Dunant et al., "Stevens-Johnson syndrome and toxic epidermal necrolysis: assessment of medication risks with emphasis on recently marketed drugs. The EuroSCAR-study," *Journal of Investigative Dermatology*, vol. 128, no. 1, pp. 35–44, 2008.
- [54] A. C. Rosen, Y. Balagula, D. W. Raisch et al., "Life-threatening dermatologic adverse events in oncology," *Anti-Cancer Drugs*, vol. 25, no. 2, pp. 225–234, 2014.
- [55] E. Dika, G. M. Ravaoli, P. A. Fanti et al., "Cutaneous adverse effects during ipilimumab treatment for metastatic melanoma: a prospective study," *European Journal of Dermatology*, vol. 27, no. 3, pp. 266–270, 2017.
- [56] K. L. Vivar, M. Deschaine, J. Messina et al., "Epidermal programmed cell death-ligand 1 expression in TEN associated with nivolumab therapy," *Journal of Cutaneous Pathology*, 2016.
- [57] C. Y. Lin, C. W. Wang, C. R. Hui et al., "Delayed-type hypersensitivity reactions induced by proton pump inhibitors: a clinical and in vitro T-cell reactivity study," *Allergy*, 2017.
- [58] B. Sassolas, C. Haddad, M. Mockenhaupt et al., "ALDEN, an algorithm for assessment of drug causality in Stevens-Johnson syndrome and toxic epidermal necrolysis: comparison with case-control analysis," *Clinical Pharmacology & Therapeutics*, vol. 88, no. 1, pp. 60–68, 2010.
- [59] J. C. Roujeau, J. P. Kelly, L. Naldi et al., "Medication use and the risk of Stevens-Johnson syndrome or toxic epidermal necrolysis," *The New England Journal of Medicine*, vol. 333, no. 24, pp. 1600–1608, 1995.
- [60] P. Sekula, A. Dunant, M. Mockenhaupt et al., "Comprehensive survival analysis of a cohort of patients with Stevens-Johnson syndrome and toxic epidermal necrolysis," *Journal of Investigative Dermatology*, vol. 133, no. 5, pp. 1197–1204, 2013.
- [61] S. Bastuji-Garin, N. Fouchard, M. Bertocchi, J. C. Roujeau, J. Revuz, and P. Wolkenstein, "SCORTEN: a severity-of-illness score for toxic epidermal necrolysis," *Journal of Investigative Dermatology*, vol. 115, no. 2, pp. 149–153, 2000.
- [62] S. Guegan, S. Bastuji-Garin, E. Poszepczynska-Guigne, J. C. Roujeau, and J. Revuz, "Performance of the SCORTEN during the first five days of hospitalization to predict the prognosis of epidermal necrolysis," *Journal of Investigative Dermatology*, vol. 126, no. 2, pp. 272–276, 2006.
- [63] A. Beck, K. P. Quirke, R. L. Gamelli, and M. J. Mosier, "Pediatric toxic epidermal necrolysis: using SCORTEN and predictive models to predict morbidity when a focus on mortality is not enough," *Journal of Burn Care & Research*, vol. 36, no. 1, pp. 167–177, 2015.
- [64] A. Sidoroff, S. Halevy, J. N. Bavinck, L. Vaillant, and J. C. Roujeau, "Acute generalized exanthematous pustulosis

- (AGEP) - a clinical reaction pattern," *Journal of Cutaneous Pathology*, vol. 28, no. 3, pp. 113–119, 2001.
- [65] E. H. Saissi, F. Beau-Salinas, A. P. Jonville-Bera, G. Lorette, E. Autret-Leca, and Centres Régionaux de Pharmacovigilance, "Drugs associated with acute generalized exanthematic pustulosis," *Annales de Dermatologie et de Venereologie*, vol. 130, no. 6-7, pp. 612–618, 2003.
- [66] J. L. Gueant, A. Romano, J. A. Cornejo-Garcia et al., "HLA-DRA variants predict penicillin allergy in genome-wide fine-mapping genotyping," *The Journal of Allergy and Clinical Immunology*, vol. 135, no. 1, pp. 253–259.e10, 2015.
- [67] A. J. Apter, H. Schelleman, A. Walker, K. Addya, and T. Rebbeck, "Clinical and genetic risk factors of self-reported penicillin allergy," *The Journal of Allergy and Clinical Immunology*, vol. 122, no. 1, pp. 152–158, 2008.
- [68] R. M. Gueant-Rodriguez, A. Romano, M. Beri-Dexheimer, M. Viola, F. Gaeta, and J. L. Gueant, "Gene-gene interactions of IL13 and IL4RA variants in immediate allergic reactions to betalactam antibiotics," *Pharmacogenetics and Genomics*, vol. 16, no. 10, pp. 713–719, 2006.
- [69] H. L. Qiao, J. Yang, and Y. W. Zhang, "Relationships between specific serum IgE, cytokines and polymorphisms in the IL-4, IL-4R α in patients with penicillins allergy," *Allergy*, vol. 60, no. 8, pp. 1053–1059, 2005.
- [70] J. Yang, H. L. Qiao, and Z. M. Dong, "Polymorphisms of IL-13 and IL-4-IL-13-SNPs in patients with penicillin allergies," *European Journal of Clinical Pharmacology*, vol. 61, no. 11, pp. 803–809, 2005.
- [71] L. Ming, Q. Wen, H. L. Qiao, and Z. M. Dong, "Interleukin-18 and IL18-607A/C and -137G/C gene polymorphisms in patients with penicillin allergy," *Journal of International Medical Research*, vol. 39, no. 2, pp. 388–398, 2011.
- [72] A. Oussalah, C. Mayorga, M. Blanca et al., "Genetic variants associated with drugs-induced immediate hypersensitivity reactions: a PRISMA-compliant systematic review," *Allergy*, vol. 71, no. 4, pp. 443–462, 2016.
- [73] A. C. Bursztejn, A. Romano, R. M. Gueant-Rodriguez et al., "Allergy to betalactams and nucleotide-binding oligomerization domain (NOD) gene polymorphisms," *Allergy*, vol. 68, no. 8, pp. 1076–1080, 2013.
- [74] S. H. Kim, Y. M. Ye, S. K. Lee, and H. S. Park, "Genetic mechanism of aspirin-induced urticaria/angioedema," *Current Opinion in Allergy and Clinical Immunology*, vol. 6, no. 4, pp. 266–270, 2006.
- [75] M. L. Pacor, G. Di Lorenzo, P. Mansueto et al., "Relationship between human leucocyte antigen class I and class II and chronic idiopathic urticaria associated with aspirin and/or NSAIDs hypersensitivity," *Mediators of Inflammation*, vol. 2006, Article ID 62489, 5 pages, 2006.
- [76] N. Palikhe, S. H. Kim, E. M. Yang et al., "Analysis of high-affinity IgE receptor (Fc ϵ R1) polymorphisms in patients with aspirin-intolerant chronic urticaria," *Allergy and Asthma Proceedings*, vol. 29, no. 3, pp. 250–257, 2008.
- [77] S. H. Kim, E. M. Yang, H. J. Park, Y. M. Ye, H. Y. Lee, and H. S. Park, "Differential contribution of the CysLTR1 gene in patients with aspirin hypersensitivity," *Journal of Clinical Immunology*, vol. 27, no. 6, pp. 613–619, 2007.
- [78] J. A. Cornejo-Garcia, L. R. Jagemann, N. Blanca-Lopez et al., "Genetic variants of the arachidonic acid pathway in non-steroidal anti-inflammatory drug-induced acute urticaria," *Clinical & Experimental Allergy*, vol. 42, no. 12, pp. 1772–1781, 2012.
- [79] P. Ayuso, M. Blanca, J. A. Cornejo-Garcia et al., "Variability in histamine receptor genes *HRH1*, *HRH2* and *HRH4* in patients with hypersensitivity to NSAIDs," *Pharmacogenomics*, vol. 14, no. 15, pp. 1871–1878, 2013.
- [80] R. Y. Pan, R. L. Dao, S. I. Hung, and W. H. Chung, "Pharmacogenomic advances in the prediction and prevention of cutaneous idiosyncratic drug reactions," *Clinical Pharmacology & Therapeutics*, vol. 102, no. 1, pp. 86–97, 2017.
- [81] S. I. Hung, W. H. Chung, Z. S. Liu et al., "Common risk allele in aromatic antiepileptic-drug induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Han Chinese," *Pharmacogenomics*, vol. 11, no. 3, pp. 349–356, 2010.
- [82] C. B. Man, P. Kwan, L. Baum et al., "Association between HLA-B*1502 allele and antiepileptic drug-induced cutaneous reactions in Han Chinese," *Epilepsia*, vol. 48, no. 5, pp. 1015–1018, 2007.
- [83] C. Lochareonkul, J. Loplumlert, C. Limotai et al., "Carbamazepine and phenytoin induced Stevens-Johnson syndrome is associated with HLA-B*1502 allele in Thai population," *Epilepsia*, vol. 49, no. 12, pp. 2087–2091, 2008.
- [84] C. B. Chen, Y. H. Hsiao, T. Wu et al., "Risk and association of HLA with oxcarbazepine-induced cutaneous adverse reactions in Asians," *Neurology*, vol. 88, no. 1, pp. 78–86, 2017.
- [85] Y. W. Shi, F. L. Min, X. R. Liu et al., "Hla-B alleles and lamotrigine-induced cutaneous adverse drug reactions in the Han Chinese population," *Basic & Clinical Pharmacology & Toxicology*, vol. 109, no. 1, pp. 42–46, 2011.
- [86] S. I. Hung, W. H. Chung, S. H. Jee et al., "Genetic susceptibility to carbamazepine-induced cutaneous adverse drug reactions," *Pharmacogenetics and Genomics*, vol. 16, no. 4, pp. 297–306, 2006.
- [87] E. Genin, D. P. Chen, S. I. Hung et al., "HLA-A*31:01 and different types of carbamazepine-induced severe cutaneous adverse reactions: an international study and meta-analysis," *The Pharmacogenomics Journal*, vol. 14, no. 3, pp. 281–288, 2014.
- [88] T. Ozeki, T. Mushiroda, A. Yowang et al., "Genome-wide association study identifies HLA-A*3101 allele as a genetic risk factor for carbamazepine-induced cutaneous adverse drug reactions in Japanese population," *Human Molecular Genetics*, vol. 20, no. 5, pp. 1034–1041, 2011.
- [89] S. H. Kim, K. W. Lee, W. J. Song et al., "Carbamazepine-induced severe cutaneous adverse reactions and HLA genotypes in Koreans," *Epilepsy Research*, vol. 97, no. 1-2, pp. 190–197, 2011.
- [90] M. McCormack, A. Alfirevic, S. Bourgeois et al., "HLA-A*3101 and carbamazepine-induced hypersensitivity reactions in Europeans," *The New England Journal of Medicine*, vol. 364, no. 12, pp. 1134–1143, 2011.
- [91] W. H. Chung, W. C. Chang, Y. S. Lee et al., "Genetic variants associated with phenytoin-related severe cutaneous adverse reactions," *JAMA*, vol. 312, no. 5, pp. 525–534, 2014.
- [92] S. I. Hung, W. H. Chung, L. B. Liou et al., "HLA-B*5801 allele as a genetic marker for severe cutaneous adverse reactions caused by allopurinol," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 11, pp. 4134–4139, 2005.

- [93] C. Lonjou, N. Borot, P. Sekula et al., "A European study of HLA-B in Stevens-Johnson syndrome and toxic epidermal necrolysis related to five high-risk drugs," *Pharmacogenetics and Genomics*, vol. 18, no. 2, pp. 99–107, 2008.
- [94] N. Kaniwa, Y. Saito, M. Aihara et al., "HLA-B locus in Japanese patients with anti-epileptics and allopurinol-related Stevens-Johnson syndrome and toxic epidermal necrolysis," *Pharmacogenomics*, vol. 9, no. 11, pp. 1617–1622, 2008.
- [95] W. Tassaneeyakul, T. Jantararoungtong, P. Chen et al., "Strong association between HLA-B*5801 and allopurinol-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in a Thai population," *Pharmacogenetics and Genomics*, vol. 19, no. 9, pp. 704–709, 2009.
- [96] H. R. Kang, Y. K. Jee, Y. S. Kim et al., "Positive and negative associations of HLA class I alleles with allopurinol-induced SCARs in Koreans," *Pharmacogenetics and Genomics*, vol. 21, no. 5, pp. 303–307, 2011.
- [97] C. Y. Ng, Y. T. Yeh, C. W. Wang et al., "Impact of the HLA-B*58:01 allele and renal impairment on allopurinol-induced cutaneous adverse reactions," *Journal of Investigative Dermatology*, vol. 136, no. 7, pp. 1373–1381, 2016.
- [98] S. Hetherington, A. R. Hughes, M. Mosteller et al., "Genetic variations in HLA-B region and hypersensitivity reactions to abacavir," *The Lancet*, vol. 359, no. 9312, pp. 1121–1122, 2002.
- [99] S. Mallal, D. Nolan, C. Witt et al., "Association between presence of HLA-B*5701, HLA-DR7, and HLA-DQ3 and hypersensitivity to HIV-1 reverse-transcriptase inhibitor abacavir," *The Lancet*, vol. 359, no. 9308, pp. 727–732, 2002.
- [100] F. A. Shepherd, J. Rodrigues Pereira, T. Ciuleanu et al., "Erlotinib in previously treated non-small-cell lung cancer," *The New England Journal of Medicine*, vol. 353, no. 2, pp. 123–132, 2005.
- [101] S. Chantarangsu, T. Mushiroda, S. Mahasirimongkol et al., "HLA-B*3505 allele is a strong predictor for nevirapine-induced skin adverse drug reactions in HIV-infected Thai patients," *Pharmacogenetics and Genomics*, vol. 19, no. 2, pp. 139–146, 2009.
- [102] H. Gatanaga, H. Yazaki, J. Tanuma et al., "HLA-Cw8 primarily associated with hypersensitivity to nevirapine," *AIDS*, vol. 21, no. 2, pp. 264–265, 2007.
- [103] B. Schnyder and W. J. Pichler, "Allergy to sulfonamides," *The Journal of Allergy and Clinical Immunology*, vol. 131, no. 1, pp. 256–257.e5, 2013.
- [104] Y. F. Lin, C. H. Yang, H. Sindy et al., "Severe cutaneous adverse reactions related to systemic antibiotics," *Clinical Infectious Diseases*, vol. 58, no. 10, pp. 1377–1385, 2014.
- [105] F. R. Zhang, H. Liu, A. Irwanto et al., "HLA-B*13:01 and the dapsone hypersensitivity syndrome," *The New England Journal of Medicine*, vol. 369, no. 17, pp. 1620–1628, 2013.
- [106] M. M. Monshi, L. Faulkner, A. Gibson et al., "Human leukocyte antigen (HLA)-B*57:01-restricted activation of drug-specific T cells provides the immunological basis for flucloxacillin-induced liver injury," *Hepatology*, vol. 57, no. 2, pp. 727–739, 2013.
- [107] M. I. Lucena, M. Molokhia, Y. Shen et al., "Susceptibility to amoxicillin-clavulanate-induced liver injury is influenced by multiple HLA class I and II alleles," *Gastroenterology*, vol. 141, no. 1, pp. 338–347, 2011.
- [108] S. H. Kim, M. Kim, K. W. Lee et al., "HLA-B*5901 is strongly associated with methazolamide-induced Stevens-Johnson syndrome/toxic epidermal necrolysis," *Pharmacogenomics*, vol. 11, no. 6, pp. 879–884, 2010.
- [109] A. K. Daly, G. P. Aithal, J. B. Leathart, R. A. Swainsbury, T. S. Dang, and C. P. Day, "Genetic susceptibility to diclofenac-induced hepatotoxicity: contribution of UGT2B7, CYP2C8, and ABC2 genotypes," *Gastroenterology*, vol. 132, no. 1, pp. 272–281, 2007.
- [110] A. K. Daly and C. P. Day, "Genetic association studies in drug-induced liver injury," *Drug Metabolism Reviews*, vol. 44, no. 1, pp. 116–126, 2012.
- [111] E. Padovan, T. Bauer, M. M. Tongio, H. Kalbacher, and H. U. Weltzien, "Penicilloyl peptides are recognized as T cell antigenic determinants in penicillin allergy," *European Journal of Immunology*, vol. 27, no. 6, pp. 1303–1307, 1997.
- [112] C. Y. Wei, W. H. Chung, H. W. Huang, Y. T. Chen, and S. I. Hung, "Direct interaction between HLA-B and carbamazepine activates T cells in patients with Stevens-Johnson syndrome," *The Journal of Allergy and Clinical Immunology*, vol. 129, no. 6, pp. 1562–1569.e5, 2012.
- [113] J. Yun, M. J. Marcaida, K. K. Eriksson et al., "Oxypurinol directly and immediately activates the drug-specific T cells via the preferential use of HLA-B*58:01," *The Journal of Immunology*, vol. 192, no. 7, pp. 2984–2993, 2014.
- [114] P. T. Illing, J. P. Vivian, N. L. Dudek et al., "Immune self-reactivity triggered by drug-modified HLA-peptide repertoire," *Nature*, vol. 486, no. 7404, pp. 554–558, 2012.
- [115] D. A. Ostrov, B. J. Grant, Y. A. Pompeu et al., "Drug hypersensitivity caused by alteration of the MHC-presented self-peptide repertoire," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 25, pp. 9959–9964, 2012.
- [116] K. D. White, W. H. Chung, S. I. Hung, S. Mallal, and E. J. Phillips, "Evolving models of the immunopathogenesis of T cell-mediated drug allergy: the role of host, pathogens, and drug response," *The Journal of Allergy and Clinical Immunology*, vol. 136, no. 2, pp. 219–234, 2015.
- [117] S. Watkins and W. J. Pichler, "Sulfamethoxazole induces a switch mechanism in T cell receptors containing TCR β 20-1, altering pHLA recognition," *PLoS One*, vol. 8, no. 10, article e76211, 2013.
- [118] K. W. Williams and H. P. Sharma, "Anaphylaxis and urticaria," *Immunology and Allergy Clinics of North America*, vol. 35, no. 1, pp. 199–219, 2015.
- [119] D. MacGlashan Jr, "Expression of CD203c and CD63 in human basophils: relationship to differential regulation of piecemeal and anaphylactic degranulation processes," *Clinical & Experimental Allergy*, vol. 40, no. 9, pp. 1365–1377, 2010.
- [120] D. W. MacGlashan Jr, "Basophil activation testing," *The Journal of Allergy and Clinical Immunology*, vol. 132, no. 4, pp. 777–787, 2013.
- [121] P. Vadas, M. Gold, B. Perelman et al., "Platelet-activating factor, PAF acetylhydrolase, and severe anaphylaxis," *The New England Journal of Medicine*, vol. 358, no. 1, pp. 28–35, 2008.
- [122] J. van der Heijden, J. Geissler, E. van Mirre et al., "A novel splice variant of Fc γ RIIa: a risk factor for anaphylaxis in patients with hypogammaglobulinemia," *The Journal*

- of *Allergy and Clinical Immunology*, vol. 131, no. 5, pp. 1408–1416.e5, 2013.
- [123] R. R. Vassallo, “Review: IgA anaphylactic transfusion reactions. Part I. Laboratory diagnosis, incidence, and supply of IgA-deficient products,” *Immunohematology*, vol. 20, no. 4, pp. 226–233, 2004.
- [124] C. Steenholdt, M. Svenson, K. Bendtzen, O. O. Thomsen, J. Brynskov, and M. A. Ainsworth, “Acute and delayed hypersensitivity reactions to infliximab and adalimumab in a patient with Crohn’s disease,” *Journal of Crohn’s and Colitis*, vol. 6, no. 1, pp. 108–111, 2012.
- [125] F. Baert, M. Noman, S. Vermeire et al., “Influence of immunogenicity on the long-term efficacy of infliximab in Crohn’s disease,” *The New England Journal of Medicine*, vol. 348, no. 7, pp. 601–608, 2003.
- [126] J. Szebeni, “Complement activation-related pseudoallergy: a stress reaction in blood triggered by nanomedicines and biologicals,” *Molecular Immunology*, vol. 61, no. 2, pp. 163–173, 2014.
- [127] T. K. Kishimoto, K. Viswanathan, T. Ganguly et al., “Contaminated heparin associated with adverse clinical events and activation of the contact system,” *The New England Journal of Medicine*, vol. 358, no. 23, pp. 2457–2467, 2008.
- [128] M. Veien, F. Szlam, J. T. Holden, K. Yamaguchi, D. D. Denson, and J. H. Levy, “Mechanisms of nonimmunological histamine and tryptase release from human cutaneous mast cells,” *Anesthesiology*, vol. 92, no. 4, pp. 1074–1081, 2000.
- [129] J. A. Blunk, M. Schmelz, S. Zeck, P. Skov, R. Likar, and W. Koppert, “Opioid-induced mast cell activation and vascular responses is not mediated by μ -opioid receptors: an *in vivo* microdialysis study in human skin,” *Anesthesia & Analgesia*, vol. 98, no. 2, pp. 364–370, 2004.
- [130] B. D. McNeil, P. Pundir, S. Meeker et al., “Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions,” *Nature*, vol. 519, no. 7542, pp. 237–241, 2015.
- [131] S. J. Posadas, A. Padial, M. J. Torres et al., “Delayed reactions to drugs show levels of perforin, granzyme B, and Fas-L to be related to disease severity,” *The Journal of Allergy and Clinical Immunology*, vol. 109, no. 1, pp. 155–161, 2002.
- [132] I. Viard, P. Wehrli, R. Bullani et al., “Inhibition of toxic epidermal necrolysis by blockade of CD95 with human intravenous immunoglobulin,” *Science*, vol. 282, no. 5388, pp. 490–493, 1998.
- [133] A. Nassif, A. Bensussan, G. Dorothee et al., “Drug specific cytotoxic T-cells in the skin lesions of a patient with toxic epidermal necrolysis,” *Journal of Investigative Dermatology*, vol. 118, no. 4, pp. 728–733, 2002.
- [134] I. Voskoboinik, J. C. Whisstock, and J. A. Trapani, “Perforin and granzymes: function, dysfunction and human pathology,” *Nature Reviews Immunology*, vol. 15, no. 6, pp. 388–400, 2015.
- [135] W. H. Chung, S. I. Hung, J. Y. Yang et al., “Granulysin is a key mediator for disseminated keratinocyte death in Stevens-Johnson syndrome and toxic epidermal necrolysis,” *Nature Medicine*, vol. 14, no. 12, pp. 1343–1350, 2008.
- [136] R. Abe, N. Yoshioka, J. Murata, Y. Fujita, and H. Shimizu, “Granulysin as a marker for early diagnosis of the Stevens-Johnson syndrome,” *Annals of Internal Medicine*, vol. 151, no. 7, pp. 514–515, 2009.
- [137] M. Weinborn, A. Barbaud, F. Truchetet, P. Beurey, L. Germain, and B. Cribier, “Histopathological study of six types of adverse cutaneous drug reactions using granulysin expression,” *International Journal of Dermatology*, vol. 55, no. 11, pp. 1225–1233, 2016.
- [138] S. SC, M. Mockenhaupt, P. Wolkenstein et al., “Interleukin-15 is associated with severity and mortality in Stevens-Johnson syndrome/toxic epidermal necrolysis,” *Journal of Investigative Dermatology*, vol. 137, pp. 1065–1073, 2017.
- [139] Z. G. Liu, “Molecular mechanism of TNF signaling and beyond,” *Cell Research*, vol. 15, no. 1, pp. 24–27, 2005.
- [140] P. Paquet, A. Nikkels, J. E. Arrese, A. Vanderkelen, and G. E. Pierard, “Macrophages and tumor necrosis factor a in toxic epidermal necrolysis,” *Archives of Dermatology*, vol. 130, no. 5, pp. 605–608, 1994.
- [141] C. Paul, P. Wolkenstein, H. Adle et al., “Apoptosis as a mechanism of keratinocyte death in toxic epidermal necrolysis,” *British Journal of Dermatology*, vol. 134, no. 4, pp. 710–714, 1996.
- [142] A. Nassif, H. Moslehi, S. Le Gouvello et al., “Evaluation of the potential role of cytokines in toxic epidermal necrolysis,” *Journal of Investigative Dermatology*, vol. 123, no. 5, pp. 850–855, 2004.
- [143] M. Caproni, D. Torchia, E. Schincaglia et al., “Expression of cytokines and chemokine receptors in the cutaneous lesions of erythema multiforme and Stevens-Johnson syndrome/toxic epidermal necrolysis,” *British Journal of Dermatology*, vol. 155, no. 4, pp. 722–728, 2006.
- [144] S. Halevy, “Acute generalized exanthematous pustulosis,” *Current Opinion in Allergy and Clinical Immunology*, vol. 9, no. 4, pp. 322–328, 2009.
- [145] P. Schaerli, M. Britschgi, M. Keller et al., “Characterization of human T cells that regulate neutrophilic skin inflammation,” *The Journal of Immunology*, vol. 173, no. 3, pp. 2151–2158, 2004.
- [146] M. Britschgi and W. J. Pichler, “Acute generalized exanthematous pustulosis, a clue to neutrophil-mediated inflammatory processes orchestrated by T cells,” *Current Opinion in Allergy and Clinical Immunology*, vol. 2, no. 4, pp. 325–331, 2002.
- [147] A. A. Navarini, L. Valeyrie-Allanore, N. Setta-Kaffetzki et al., “Rare variations in *IL36RN* in severe adverse drug reactions manifesting as acute generalized exanthematous pustulosis,” *Journal of Investigative Dermatology*, vol. 133, no. 7, pp. 1904–1907, 2013.
- [148] H. S. Song, S. J. Kim, T. I. Park, Y. H. Jang, and E. S. Lee, “Immunohistochemical comparison of IL-36 and the IL-23/Th17 axis of generalized pustular psoriasis and acute generalized exanthematous pustulosis,” *Annals of Dermatology*, vol. 28, no. 4, pp. 451–456, 2016.
- [149] S. A. Walsh and D. Creamer, “Drug reaction with eosinophilia and systemic symptoms (DRESS): a clinical update and review of current thinking,” *Clinical and Experimental Dermatology*, vol. 36, no. 1, pp. 6–11, 2011.
- [150] T. Komatsu-Fujii, Y. Chinuki, H. Niihara et al., “The thymus and activation-regulated chemokine (TARC) level in serum at an early stage of a drug eruption is a prognostic biomarker of severity of systemic inflammation,” *Allergy International*, 2017.
- [151] K. Ogawa, H. Morito, A. Hasegawa et al., “Identification of thymus and activation-regulated chemokine (TARC/CCL17)

- as a potential marker for early indication of disease and prediction of disease activity in drug-induced hypersensitivity syndrome (DIHS)/drug rash with eosinophilia and systemic symptoms (DRESS)," *Journal of Dermatological Science*, vol. 69, no. 1, pp. 38–43, 2013.
- [152] B. Tapia, A. Padiar, E. Sanchez-Sabate et al., "Involvement of CCL27-CCR10 interactions in drug-induced cutaneous reactions," *The Journal of Allergy and Clinical Immunology*, vol. 114, no. 2, pp. 335–340, 2004.
- [153] O. Correia, L. Delgado, I. L. Barbosa, F. Campilho, and J. Fleming-Torrinha, "Increased interleukin 10, tumor necrosis factor α , and interleukin 6 levels in blister fluid of toxic epidermal necrolysis," *Journal of the American Academy of Dermatology*, vol. 47, no. 1, pp. 58–62, 2002.
- [154] P. Paquet, F. Paquet, W. Al Saleh, P. Reper, A. Vanderkelen, and G. E. Pierard, "Immunoregulatory effector cells in drug-induced toxic epidermal necrolysis," *The American Journal of Dermatopathology*, vol. 22, no. 5, pp. 413–417, 2000.
- [155] W. H. Chung, W. C. Chang, S. L. Stocker et al., "Insights into the poor prognosis of allopurinol-induced severe cutaneous adverse reactions: the impact of renal insufficiency, high plasma levels of oxypurinol and granulysin," *Annals of the Rheumatic Diseases*, vol. 74, no. 12, pp. 2157–2164, 2015.
- [156] R. Takahashi, Y. Kano, Y. Yamazaki, M. Kimishima, Y. Mizukawa, and T. Shiohara, "Defective regulatory T cells in patients with severe drug eruptions: timing of the dysfunction is associated with the pathological phenotype and outcome," *The Journal of Immunology*, vol. 182, no. 12, pp. 8071–8079, 2009.
- [157] T. Shiohara, M. Inaoka, and Y. Kano, "Drug-induced hypersensitivity syndrome (DIHS): a reaction induced by a complex interplay among herpesviruses and antiviral and antidrug immune responses," *Allergology International*, vol. 55, no. 1, pp. 1–8, 2006.
- [158] T. Shiohara, M. Kurata, Y. Mizukawa, and Y. Kano, "Recognition of immune reconstitution syndrome necessary for better management of patients with severe drug eruptions and those under immunosuppressive therapy," *Allergology International*, vol. 59, no. 4, pp. 333–343, 2010.
- [159] T. Shiohara, Y. Ushigome, Y. Kano, and R. Takahashi, "Crucial role of viral reactivation in the development of severe drug eruptions: a comprehensive review," *Clinical Reviews in Allergy & Immunology*, vol. 49, no. 2, pp. 192–202, 2015.
- [160] T. Yoshikawa, A. Fujita, A. Yagami et al., "Human herpesvirus 6 reactivation and inflammatory cytokine production in patients with drug-induced hypersensitivity syndrome," *Journal of Clinical Virology*, vol. 37, Supplement 1, pp. S92–S96, 2006.
- [161] T. Shiohara and Y. Kano, "A complex interaction between drug allergy and viral infection," *Clinical Reviews in Allergy & Immunology*, vol. 33, no. 1–2, pp. 124–133, 2007.
- [162] F. E. Simons, L. R. Arduzzo, M. B. Biló et al., "2012 update: World Allergy Organization guidelines for the assessment and management of anaphylaxis," *Current Opinion in Allergy and Clinical Immunology*, vol. 12, no. 4, pp. 389–399, 2012.
- [163] R. Y. Lin, L. B. Schwartz, A. Curry et al., "Histamine and tryptase levels in patients with acute allergic reactions: an emergency department-based study," *The Journal of Allergy and Clinical Immunology*, vol. 106, no. 1, pp. 65–71, 2000.
- [164] F. E. Simons, A. J. Frew, I. J. Ansotegui et al., "Risk assessment in anaphylaxis: current and future approaches," *The Journal of Allergy and Clinical Immunology*, vol. 120, no. 1, Supplement, pp. S2–S24, 2007.
- [165] E. Gomez, M. J. Torres, C. Mayorga, and M. Blanca, "Immunologic evaluation of drug allergy," *Allergy, Asthma & Immunology Research*, vol. 4, no. 5, pp. 251–263, 2012.
- [166] M. Blanca, C. Mayorga, M. J. Torres et al., "Clinical evaluation of Pharmacia CAP system™ RAST FEIA amoxicilloyl and benzylpenicilloyl in patients with penicillin allergy," *Allergy*, vol. 56, no. 9, pp. 862–870, 2001.
- [167] I. Dona, M. J. Torres, M. I. Montanez, and T. D. Fernandez, "In vitro diagnostic testing for antibiotic allergy," *Allergy, Asthma & Immunology Research*, vol. 9, no. 4, pp. 288–298, 2017.
- [168] C. Mayorga, I. Dona, E. Perez-Inestrosa, T. D. Fernandez, and M. J. Torres, "The value of in vitro tests to diminish drug challenges," *International Journal of Molecular Sciences*, vol. 18, no. 6, 2017.
- [169] C. Mayorga, G. Celik, P. Rouzaire et al., "In vitro tests for drug hypersensitivity reactions: an ENDA/EAACI Drug Allergy Interest Group position paper," *Allergy*, vol. 71, no. 8, pp. 1103–1134, 2016.
- [170] H. J. Hoffmann, A. F. Santos, C. Mayorga et al., "The clinical utility of basophil activation testing in diagnosis and monitoring of allergic disease," *Allergy*, vol. 70, no. 11, pp. 1393–1405, 2015.
- [171] J. Leysen, V. Sabato, M. M. Verweij et al., "The basophil activation test in the diagnosis of immediate drug hypersensitivity," *Expert Reviews of Clinical Immunology*, vol. 7, no. 3, pp. 349–355, 2011.
- [172] E. C. McGowan and S. Saini, "Update on the performance and application of basophil activation tests," *Current Allergy and Asthma Reports*, vol. 13, no. 1, pp. 101–109, 2013.
- [173] A. L. De Week, M. L. Sanz, P. M. Gamboa et al., "Diagnosis of immediate-type β -lactam allergy in vitro by flow-cytometric basophil activation test and sulfidoleukotriene production: a multicenter study," *Journal of Investigational Allergology & Clinical Immunology*, vol. 19, no. 2, pp. 91–109, 2009.
- [174] N. Saito, R. Abe, N. Yoshioka, J. Murata, Y. Fujita, and H. Shimizu, "Prolonged elevation of serum granulysin in drug-induced hypersensitivity syndrome," *British Journal of Dermatology*, vol. 167, no. 2, pp. 452–453, 2012.
- [175] L. Feldmeyer, K. Heidemeyer, and N. Yawalkar, "Acute generalized exanthematous pustulosis: pathogenesis, genetic background, clinical variants and therapy," *International Journal of Molecular Sciences*, vol. 17, no. 8, 2016.
- [176] S. SC, S. I. Hung, W. L. Fan, R. L. Dao, and W. H. Chung, "Severe cutaneous adverse reactions: the pharmacogenomics from research to clinical implementation," *International Journal of Molecular Sciences*, vol. 17, p. 1890, 2016.
- [177] A. A. Elzagallaai and M. J. Rieder, "In vitro testing for diagnosis of idiosyncratic adverse drug reactions: implications for pathophysiology," *British Journal of Clinical Pharmacology*, vol. 80, no. 4, pp. 889–900, 2015.
- [178] C. M. Rive, J. Bourke, and E. J. Phillips, "Testing for drug hypersensitivity syndromes," *The Clinical Biochemist Reviews*, vol. 34, no. 1, pp. 15–38, 2013.
- [179] G. Porebski, "In vitro assays in severe cutaneous adverse drug reactions: are they still research tools or diagnostic tests already?," *International Journal of Molecular Sciences*, vol. 18, no. 8, p. 1737, 2017.

- [180] W. J. Pichler and J. Tilch, "The lymphocyte transformation test in the diagnosis of drug hypersensitivity," *Allergy*, vol. 59, no. 8, pp. 809–820, 2004.
- [181] A. T. Nagao-Dias, F. M. Teixeira, and H. L. Coelho, "Diagnosing immune-mediated reactions to drugs," *Allergologia et Immunopathologia*, vol. 37, no. 2, pp. 98–104, 2009.
- [182] Y. Kano, K. Hirahara, Y. Mitsuyama, R. Takahashi, and T. Shiohara, "Utility of the lymphocyte transformation test in the diagnosis of drug sensitivity: dependence on its timing and the type of drug eruption," *Allergy*, vol. 62, no. 12, pp. 1439–1444, 2007.
- [183] Y. Srinoulprasert and W. J. Pichler, "Enhancement of drug-specific lymphocyte proliferation using CD25^{hi}-depleted CD3⁺ effector cells," *International Archives of Allergy and Immunology*, vol. 163, no. 3, pp. 198–205, 2014.
- [184] K. Kato, A. Kawase, H. Azukizawa et al., "Novel interferon- γ enzyme-linked immunospot assay using activated cells for identifying hypersensitivity-inducing drug culprits," *Journal of Dermatological Science*, vol. 86, no. 3, pp. 222–229, 2017.
- [185] L. Valeyrie-Allanore, M. Mockenhaupt, P. Sekula et al., "Mechanisms that limit proliferative potential of drug-specific LTT in drug-induced severe cutaneous adverse reaction patients," *Clinical and Translational Allergy*, vol. 4, Supplement 3, pp. O1–P149, 2014.
- [186] W. H. Chung, R. Y. Pan, M. T. Chu et al., "Oxypurinol-specific T cells possess preferential TCR clonotypes and express granulysin in allopurinol-induced severe cutaneous adverse reactions," *Journal of Investigative Dermatology*, vol. 135, no. 9, pp. 2237–2248, 2015.
- [187] A. Barbaud, E. Collet, B. Milpied et al., "A multicentre study to determine the value and safety of drug patch tests for the three main classes of severe cutaneous adverse drug reactions," *British Journal of Dermatology*, vol. 168, no. 3, pp. 555–562, 2013.
- [188] Y. T. Lin, Y. C. Chang, R. C. Hui et al., "A patch testing and cross-sensitivity study of carbamazepine-induced severe cutaneous adverse drug reactions," *Journal of the European Academy of Dermatology and Venereology*, vol. 27, no. 3, pp. 356–364, 2013.
- [189] M. J. Torres, A. Romano, G. Celik et al., "Approach to the diagnosis of drug hypersensitivity reactions: similarities and differences between Europe and North America," *Clinical and Translational Allergy*, vol. 7, no. 1, p. 7, 2017.
- [190] Joint Task Force on Practice Parameters, American Academy of Allergy, Asthma and Immunology et al., "Drug allergy: an updated practice parameter," *Annals of Allergy, Asthma & Immunology*, vol. 105, no. 4, pp. 259–273.e78, 2010.
- [191] K. Brockow, A. Romano, M. Blanca, J. Ring, W. Pichler, and P. Demoly, "General considerations for skin test procedures in the diagnosis of drug hypersensitivity," *Allergy*, vol. 57, no. 1, pp. 45–51, 2002.
- [192] T. A. Duong, L. Valeyrie-Allanore, P. Wolkenstein, and O. Chosidow, "Severe cutaneous adverse reactions to drugs," *The Lancet*, vol. 390, no. 10106, pp. 1996–2011, 2017.
- [193] M. Alrasbi and A. Sheikh, "Comparison of international guidelines for the emergency medical management of anaphylaxis," *Allergy*, vol. 62, no. 8, pp. 838–841, 2007.
- [194] A. Sheikh, Y. A. Shehata, S. G. Brown, and F. E. Simons, "Adrenaline for the treatment of anaphylaxis: Cochrane systematic review," *Allergy*, vol. 64, no. 2, pp. 204–212, 2009.
- [195] T. Kawano, F. X. Scheuermeyer, R. Stenstrom, B. H. Rowe, E. Grafstein, and B. Grunau, "Epinephrine use in older patients with anaphylaxis: clinical outcomes and cardiovascular complications," *Resuscitation*, vol. 112, pp. 53–58, 2017.
- [196] M. Thomas and I. Crawford, "Best evidence topic report. Glucagon infusion in refractory anaphylactic shock in patients on beta-blockers," *Emergency Medicine Journal*, vol. 22, no. 4, pp. 272–273, 2005.
- [197] K. J. Choo, F. E. Simons, and A. Sheikh, "Glucocorticoids for the treatment of anaphylaxis," *Cochrane Database of Systematic Reviews*, no. 4, article CD007596, 2012.
- [198] K. A. Michelson, M. C. Monuteaux, and M. I. Neuman, "Glucocorticoids and hospital length of stay for children with anaphylaxis: a retrospective study," *The Journal of Pediatrics*, vol. 167, no. 3, pp. 719–724.e3, 2015.
- [199] A. Sheikh, V. Ten Broek, S. G. Brown, and F. E. Simons, "H₁-antihistamines for the treatment of anaphylaxis: Cochrane systematic review," *Allergy*, vol. 62, no. 8, pp. 830–837, 2007.
- [200] P. Lieberman, R. A. Nicklas, C. Randolph et al., "Anaphylaxis—a practice parameter update 2015," *Annals of Allergy, Asthma & Immunology*, vol. 115, no. 5, pp. 341–384, 2015.
- [201] R. P. Dodiuk-Gad, W. H. Chung, C. H. Yang, L. CW, R. C. Hui, and N. H. Shear, "The 8th International Congress on Cutaneous Adverse Drug Reactions, Taiwan, 2013: focus on severe cutaneous adverse reactions," *Drug Safety*, vol. 37, no. 6, pp. 459–464, 2014.
- [202] H. Y. Lee, A. Dunant, P. Sekula et al., "The role of prior corticosteroid use on the clinical course of Stevens-Johnson syndrome and toxic epidermal necrolysis: a case-control analysis of patients selected from the multinational EuroSCAR and RegiSCAR studies," *British Journal of Dermatology*, vol. 167, no. 3, pp. 555–562, 2012.
- [203] E. H. Law and M. Leung, "Corticosteroids in Stevens-Johnson syndrome/toxic epidermal necrolysis: current evidence and implications for future research," *Annals of Pharmacotherapy*, vol. 49, no. 3, pp. 335–342, 2015.
- [204] Y. C. Huang, Y. C. Li, and T. J. Chen, "The efficacy of intravenous immunoglobulin for the treatment of toxic epidermal necrolysis: a systematic review and meta-analysis," *British Journal of Dermatology*, vol. 167, no. 2, pp. 424–432, 2012.
- [205] Y. C. Huang, Y. N. Chien, Y. T. Chen, Y. C. Li, and T. J. Chen, "Intravenous immunoglobulin for the treatment of toxic epidermal necrolysis: a systematic review and meta-analysis," *Giornale Italiano di Dermatologia e Venereologia*, vol. 151, no. 5, pp. 515–524, 2016.
- [206] P. Tristani-Firouzi, M. J. Petersen, J. R. Saffle, S. E. Morris, and J. J. Zone, "Treatment of toxic epidermal necrolysis with intravenous immunoglobulin in children," *Journal of the American Academy of Dermatology*, vol. 47, no. 4, pp. 548–552, 2002.
- [207] L. Valeyrie-Allanore, P. Wolkenstein, L. Brochard et al., "Open trial of ciclosporin treatment for Stevens-Johnson syndrome and toxic epidermal necrolysis," *British Journal of Dermatology*, vol. 163, no. 4, pp. 847–853, 2010.
- [208] H. Y. Lee, S. Fook-Chong, H. Y. Koh, T. Thirumoorthy, and S. M. Pang, "Cyclosporine treatment for Stevens-Johnson syndrome/toxic epidermal necrolysis: retrospective analysis of a cohort treated in a specialized referral center," *Journal of the American Academy of Dermatology*, vol. 76, no. 1, pp. 106–113, 2017.

- [209] Y.-T. H. Chen, C. Y.-N. Che-Yuan, W.-R. Lee, and Y.-C. Huang, "Efficacy of cyclosporine for the treatment of Stevens-Johnson syndrome and toxic epidermal necrolysis: systemic review and meta-analysis," *Dermatologica Sinica*, vol. 35, no. 3, pp. 131–137, 2017.
- [210] A. Wojtkiewicz, M. Wysocki, J. Fortuna, M. Chrupek, M. Matczuk, and A. Koltan, "Beneficial and rapid effect of infliximab on the course of toxic epidermal necrolysis," *Acta Dermato-Venereologica*, vol. 88, no. 4, pp. 420–421, 2008.
- [211] A. Paradisi, D. Abeni, F. Bergamo, F. Ricci, D. Didona, and B. Didona, "Etanercept therapy for toxic epidermal necrolysis," *Journal of the American Academy of Dermatology*, vol. 71, no. 2, pp. 278–283, 2014.
- [212] M. Fischer, E. Fiedler, W. C. Marsch, and J. Wohlrab, "Antitumour necrosis factor- α antibodies (infliximab) in the treatment of a patient with toxic epidermal necrolysis," *British Journal of Dermatology*, vol. 146, no. 4, pp. 707–709, 2002.
- [213] R. E. Hunger, T. Hunziker, U. Buettiker, L. R. Braathen, and N. Yawalkar, "Rapid resolution of toxic epidermal necrolysis with anti-TNF- α treatment," *The Journal of Allergy and Clinical Immunology*, vol. 116, no. 4, pp. 923–924, 2005.
- [214] B. Krefit, J. Wohlrab, I. Bramsiede, R. Eismann, M. Winkler, and W. C. Marsch, "Etoricoxib-induced toxic epidermal necrolysis: successful treatment with infliximab," *The Journal of Dermatology*, vol. 37, no. 10, pp. 904–906, 2010.
- [215] L. C. Zarate-Correa, D. C. Carrillo-Gomez, A. F. Ramirez-Escobar, and C. Serrano-Reyes, "Toxic epidermal necrolysis successfully treated with infliximab," *Journal of Investigational Allergology & Clinical Immunology*, vol. 23, no. 1, pp. 61–63, 2013.
- [216] S. Zimmermann, P. Sekula, M. Venhoff et al., "Systemic immunomodulating therapies for Stevens-Johnson syndrome and toxic epidermal necrolysis: a systematic review and meta-analysis," *JAMA Dermatology*, vol. 153, no. 6, pp. 514–522, 2017.
- [217] R. S. Stern and S. J. Divito, "Stevens-Johnson syndrome and toxic epidermal necrolysis: associations, outcomes, and pathobiology—thirty years of progress but still much to be done," *Journal of Investigative Dermatology*, vol. 137, no. 5, pp. 1004–1008, 2017.
- [218] Y. T. Cho, C. W. Yang, and C. Y. Chu, "Drug reaction with eosinophilia and systemic symptoms (DRESS): an interplay among drugs, viruses, and immune system," *International Journal of Molecular Sciences*, vol. 18, no. 6, 2017.
- [219] T. Ishida, Y. Kano, Y. Mizukawa, and T. Shiohara, "The dynamics of herpesvirus reactivations during and after severe drug eruptions: their relation to the clinical phenotype and therapeutic outcome," *Allergy*, vol. 69, no. 6, pp. 798–805, 2014.
- [220] V. Descamps, B. Ben Said, B. Sassolas et al., "Management of drug reaction with eosinophilia and systemic symptoms (DRESS)," *Annales de Dermatologie et de Vénérologie*, vol. 137, no. 11, pp. 703–708, 2010.
- [221] Z. Husain, B. Y. Reddy, and R. A. Schwartz, "DRESS syndrome: part II. Management and therapeutics," *Journal of the American Academy of Dermatology*, vol. 68, no. 5, pp. 709.e1–709.e9, 2013.
- [222] P. Joly, B. Janela, F. Tetart et al., "Poor benefit/risk balance of intravenous immunoglobulins in DRESS," *Archives of Dermatology*, vol. 148, no. 4, pp. 543–544, 2012.
- [223] M. G. Kirchhof, A. Wong, and J. P. Dutz, "Cyclosporine treatment of drug-induced hypersensitivity syndrome," *JAMA Dermatology*, vol. 152, no. 11, pp. 1254–1257, 2016.
- [224] C. Hotz, L. Valeyrie-Allanore, C. Haddad et al., "Systemic involvement of acute generalized exanthematous pustulosis: a retrospective study on 58 patients," *British Journal of Dermatology*, vol. 169, no. 6, pp. 1223–1232, 2013.
- [225] S. Ingen-Housz-Oro, C. Hotz, L. Valeyrie-Allanore et al., "Acute generalized exanthematous pustulosis: a retrospective audit of practice between 1994 and 2011 at a single centre," *British Journal of Dermatology*, vol. 172, no. 5, pp. 1455–1457, 2015.
- [226] T. Y. Mehta, L. M. Prajapati, B. Mittal et al., "Association of HLA-B*1502 allele and carbamazepine-induced Stevens-Johnson syndrome among Indians," *Indian Journal of Dermatology, Venereology and Leprology*, vol. 75, no. 6, pp. 579–582, 2009.
- [227] C. C. Chang, C. L. Too, S. Murad, and S. H. Hussein, "Association of HLA-B*1502 allele with carbamazepine-induced toxic epidermal necrolysis and Stevens-Johnson syndrome in the multi-ethnic Malaysian population," *International Journal of Dermatology*, vol. 50, no. 2, pp. 221–224, 2011.
- [228] K. W. Chong, D. W. Chan, Y. B. Cheung et al., "Association of carbamazepine-induced severe cutaneous drug reactions and HLA-B*1502 allele status, and dose and treatment duration in paediatric neurology patients in Singapore," *Archives of Disease in Childhood*, vol. 99, no. 6, pp. 581–584, 2014.
- [229] D. V. Nguyen, H. C. Chu, D. V. Nguyen et al., "HLA-B*1502 and carbamazepine-induced severe cutaneous adverse drug reactions in Vietnamese," *Asia Pacific Allergy*, vol. 5, no. 2, pp. 68–77, 2015.
- [230] P. K. Kwan, M. H. Ng, and S. V. Lo, "Association between HLA-B*15:02 allele and antiepileptic drug-induced severe cutaneous reactions in Hong Kong Chinese: a population-based study," *Hong Kong Medical Journal*, no. 20, Supplement 7, pp. S16–S18, 2014.
- [231] E. Ramírez, T. Bellón, H. Y. Tong et al., "Significant HLA class I type associations with aromatic antiepileptic drug (AED)-induced SJS/TEN are different from those found for the same AED-induced DRESS in the Spanish population," *Pharmacological Research*, vol. 115, pp. 168–178, 2017.
- [232] N. Kaniwa, Y. Saito, M. Aihara et al., "HLA-B*1511 is a risk factor for carbamazepine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Japanese patients," *Epilepsia*, vol. 51, no. 12, pp. 2461–2465, 2010.
- [233] D. Sun, Y. CH, Z. S. Liu et al., "Association of HLA-B*1502 and *1511 allele with antiepileptic drug-induced Stevens-Johnson syndrome in central China," *Journal of Huazhong University of Science and Technology Medical Sciences*, vol. 34, pp. 146–150, 2014.
- [234] H. Ikeda, Y. Takahashi, E. Yamazaki et al., "HLA class I markers in Japanese patients with carbamazepine-induced cutaneous adverse reactions," *Epilepsia*, vol. 51, no. 2, pp. 297–300, 2010.
- [235] W. Tassaneeyakul, N. Prabhmechai, C. Sukasem et al., "Associations between HLA class I and cytochrome P450 2C9 genetic polymorphisms and phenytoin-related severe cutaneous adverse reactions in a Thai population," *Pharmacogenetics and Genomics*, vol. 26, no. 5, pp. 225–234, 2016.
- [236] C. C. Chang, C. C. Ng, C. L. Too et al., "Association of HLA-B*15:13 and HLA-B*15:02 with phenytoin-induced severe

- cutaneous adverse reactions in a Malay population," *The Pharmacogenomics Journal*, vol. 17, no. 2, p. 170, 2016.
- [237] T. Zeng, Y. S. Long, F. L. Min, W. P. Liao, and Y. W. Shi, "Association of HLA-B*1502 allele with lamotrigine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Han Chinese subjects: a meta-analysis," *International Journal of Dermatology*, vol. 54, no. 4, pp. 488–493, 2015.
- [238] G. R. Kazeem, C. Cox, J. Aponte et al., "High-resolution HLA genotyping and severe cutaneous adverse reactions in lamotrigine-treated patients," *Pharmacogenetics and Genomics*, vol. 19, no. 9, pp. 661–665, 2009.
- [239] B. K. Kim, J. W. Jung, T. B. Kim et al., "HLA-A*31:01 and lamotrigine-induced severe cutaneous adverse drug reactions in a Korean population," *Annals of Allergy Asthma & Immunology*, vol. 119, no. 5, pp. 629–630, 2017.
- [240] A. M. Martin, D. Nolan, I. James et al., "Predisposition to nevirapine hypersensitivity associated with HLA-DRB1*0101 and abrogated by low CD4 T-cell counts," *AIDS*, vol. 19, no. 1, pp. 97–99, 2005.
- [241] R. Littera, C. Carcassi, A. Masala et al., "HLA-dependent hypersensitivity to nevirapine in Sardinian HIV patients," *AIDS*, vol. 20, no. 12, pp. 1621–1626, 2006.
- [242] D. F. Carr, M. Chaponda, A. L. Jorgensen et al., "Association of human leukocyte antigen alleles and nevirapine hypersensitivity in a Malawian HIV-infected population," *Clinical Infectious Diseases*, vol. 56, no. 9, pp. 1330–1339, 2013.
- [243] J. Yang, H. L. Qiao, Y. W. Zhang, L. J. Jia, X. Tian, and N. Gao, "HLA-DRB genotype and specific IgE responses in patients with allergies to penicillins," *Chinese Medical Journal*, vol. 119, no. 6, pp. 458–466, 2006.
- [244] T. Kongpan, S. Mahasirimongkol, P. Konyoung et al., "Candidate HLA genes for prediction of co-trimoxazole-induced severe cutaneous reactions," *Pharmacogenetic and Genomics*, vol. 25, no. 8, pp. 402–411, 2015.
- [245] J. Revuz, D. Penso, J. C. Roujeau et al., "Toxic epidermal necrolysis. Clinical findings and prognosis factors in 87 patients," *Archives of Dermatology*, vol. 123, no. 9, pp. 1160–1165, 1987.
- [246] J. Quiralte, F. Sanchez-Garcia, M. J. Torres et al., "Association of HLA-DR11 with the anaphylactoid reaction caused by nonsteroidal anti-inflammatory drugs," *The Journal of Allergy and Clinical Immunology*, vol. 103, no. 4, pp. 685–689, 1999.
- [247] H. L. Qiao, J. Yang, and Y. W. Zhang, "Specific serum IgE levels and FcεRIβ genetic polymorphism in patients with penicillins allergy," *Allergy*, vol. 59, no. 12, pp. 1326–1332, 2004.
- [248] Y. H. Nam, J. E. Kim, S. H. Kim et al., "Identifying genetic susceptibility to sensitization to cephalosporins in health care workers," *Journal of Korean Medical Science*, vol. 27, no. 11, pp. 1292–1299, 2012.
- [249] H. L. Qiao, Q. Wen, N. Gao, X. Tian, and L. J. Jia, "Association of IL-10 level and IL-10 promoter SNPs with specific antibodies in penicillin-allergic patients," *European Journal of Clinical Pharmacology*, vol. 63, no. 3, pp. 263–269, 2007.
- [250] N. Gao, H.-L. Qiao, L.-J. Jia, X. Tian, and Y.-W. Zhang, "Relationships between specific serum IgE, IgG, IFN-γ level and IFN-γ, IFNR1 polymorphisms in patients with penicillin allergy," *European Journal of Clinical Pharmacology*, vol. 64, no. 10, pp. 971–977, 2008.
- [251] C.-Z. Huang, J. Yang, H.-L. Qiao, and L.-J. Jia, "Polymorphisms and haplotype analysis of IL-4Rα Q576R and I75V in patients with penicillin allergy," *European Journal of Clinical Pharmacology*, vol. 65, no. 9, pp. 895–902, 2009.
- [252] C. Z. Huang, D. Zou, J. Yang, and H. L. Qiao, "Polymorphisms of STAT6 and specific serum IgE levels in patients with penicillin allergy," *International Journal of Clinical Pharmacology and Therapeutics*, vol. 50, no. 07, pp. 461–467, 2012.
- [253] L. Guglielmi, C. Fontaine, C. Gougat et al., "IL-10 promoter and IL-4R α gene SNPs are associated with immediate β-lactam allergy in atopic women," *Allergy*, vol. 61, no. 8, pp. 921–927, 2006.
- [254] R. M. Gueant-Rodriguez, J. L. Gueant, M. Viola, D. Tramoy, F. Gaeta, and A. Romano, "Association of tumor necrosis factor-α -308G>A polymorphism with IgE-mediated allergy to betalactams in an Italian population," *The Pharmacogenomics Journal*, vol. 8, no. 2, pp. 162–168, 2008.
- [255] J. A. Cornejo-Garcia, R. M. Gueant-Rodriguez, M. J. Torres et al., "Biological and genetic determinants of atopy are predictors of immediate-type allergy to betalactams, in Spain," *Allergy*, vol. 67, no. 9, pp. 1181–1185, 2012.
- [256] S. H. Kim, J. H. Choi, J. W. Holloway et al., "Leukotriene-related gene polymorphisms in patients with aspirin-intolerant urticaria and aspirin-intolerant asthma: differing contributions of ALOX5 polymorphism in Korean population," *Journal of Korean Medical Science*, vol. 20, no. 6, pp. 926–931, 2005.
- [257] J.-S. Bae, S.-H. Kim, Y.-M. Ye et al., "Significant association of FcεRIα promoter polymorphisms with aspirin-intolerant chronic urticaria," *The Journal of Allergy and Clinical Immunology*, vol. 119, no. 2, pp. 449–456.
- [258] H. J. Park, Y. M. Ye, G. Y. Hur, S. H. Kim, and H. S. Park, "Association between a TGFβ1 promoter polymorphism and the phenotype of aspirin-intolerant chronic urticaria in a Korean population," *Journal of Clinical Pharmacy and Therapeutics*, vol. 33, no. 6, pp. 691–697, 2008.
- [259] J. H. Choi, S. H. Kim, B. Y. Cho et al., "Association of TNF-α promoter polymorphisms with aspirin-induced urticaria," *Journal of Clinical Pharmacy and Therapeutics*, vol. 34, no. 2, pp. 231–238, 2009.
- [260] S. H. Kim, Y. M. Kang, S. H. Kim et al., "Histamine N-methyltransferase 939A>G polymorphism affects mRNA stability in patients with acetylsalicylic acid-intolerant chronic urticaria," *Allergy*, vol. 64, pp. 213–221, 2009.
- [261] S. H. Kim, J. K. Son, E. M. Yang, J. E. Kim, and H. S. Park, "A functional promoter polymorphism of the human IL18 gene is associated with aspirin-induced urticaria," *British Journal of Dermatology*, vol. 165, no. 5, pp. 976–984, 2011.
- [262] N. S. Palikhe, S. H. Kim, H. Y. Lee, J. H. Kim, Y. M. Ye, and H. S. Park, "Association of thromboxane A2 receptor (TBXA2R) gene polymorphism in patients with aspirin-intolerant acute urticaria," *Clinical & Experimental Allergy*, vol. 41, no. 2, pp. 179–185, 2011.
- [263] N. S. Palikhe, H. J. Sin, S. H. Kim et al., "Genetic variability of prostaglandin E2 receptor subtype EP4 gene in aspirin-intolerant chronic urticaria," *Journal of Human Genetics*, vol. 57, no. 8, pp. 494–499, 2012.

- [264] L. Mastalerz, M. Setkowicz, M. Sanak, H. Rybarczyk, and A. Szczeklik, "Familial aggregation of aspirin-induced urticaria and leukotriene C₄ synthase allelic variant," *British Journal of Dermatology*, vol. 154, no. 2, pp. 256–260, 2006.
- [265] M. Sanchez-Borges, N. Acevedo, C. Vergara et al., "The A-444C polymorphism in the leukotriene C₄ synthase gene is associated with aspirin-induced urticaria," *Journal of Investigational Allergology and Clinical Immunology*, vol. 19, no. 5, pp. 375–382, 2009.
- [266] J. A. Agundez, P. Ayuso, J. A. Cornejo-Garcia et al., "The diamine oxidase gene is associated with hypersensitivity response to non-steroidal anti-inflammatory drugs," *PLoS One*, vol. 7, no. 11, article e47571, 2012.
- [267] C. Vidal, L. Porras-Hurtado, R. Cruz et al., "Association of thromboxane A1 synthase (TBXAS1) gene polymorphism with acute urticaria induced by nonsteroidal anti-inflammatory drugs," *The Journal of Allergy and Clinical Immunology*, vol. 132, no. 4, pp. 989–991, 2013.
- [268] C. Plaza-Seron Mdel, P. Ayuso, N. Perez-Sanchez et al., "Copy number variation in ALOX5 and PTGER1 is associated with NSAIDs-induced urticaria and/or angioedema," *Pharmacogenetics and Genomics*, vol. 26, no. 6, pp. 280–287, 2016.
- [269] L. M. Ferreira Vasconcelos, R. O. Rodrigues, A. A. Albuquerque et al., "Polymorphism of *IL10*, *IL4*, *CTLA4*, and *DAO* genes in cross-reactive nonsteroidal anti-inflammatory drug hypersensitivity," *The Journal of Clinical Pharmacology*, 2017.
- [270] A. Beeler, O. Engler, B. O. Gerber, and W. J. Pichler, "Long-lasting reactivity and high frequency of drug-specific T cells after severe systemic drug hypersensitivity reactions," *Journal of Allergy and Clinical Immunology*, vol. 117, no. 2, pp. 455–462, 2006.
- [271] D. Nishio, K. Izu, K. Kabashima, and Y. Tokura, "T cell populations propagating in the peripheral blood of patients with drug eruptions," *Journal of Dermatological Science*, vol. 48, no. 1, pp. 25–33, 2007.
- [272] M. Hertl, H. Bohlen, F. Jugert, C. Boecker, R. Knaup, and H. F. Merk, "Predominance of epidermal CD8+ T lymphocytes in bullous cutaneous reactions caused by β -lactam antibiotics," *Journal of Investigative Dermatology*, vol. 101, no. 6, pp. 794–799, 1993.
- [273] K. Ogawa, H. Morito, A. Hasegawa et al., "Elevated serum thymus and activation-regulated chemokine (TARC/CCL17) relates to reactivation of human herpesvirus 6 in drug reaction with eosinophilia and systemic symptoms (DRESS)/drug-induced hypersensitivity syndrome (DIHS)," *British Journal of Dermatology*, vol. 171, no. 2, pp. 425–427, 2014.
- [274] I. Viard-Leveugle, O. Gaide, D. Jankovic et al., "TNF- α and IFN- γ are potential inducers of Fas-mediated keratinocyte apoptosis through activation of inducible nitric oxide synthase in toxic epidermal necrolysis," *Journal of Investigative Dermatology*, vol. 133, no. 2, pp. 489–498, 2013.
- [275] P. Paquet and G. E. Pierard, "Erythema multiforme and toxic epidermal necrolysis: a comparative study," *The American Journal of Dermatopathology*, vol. 19, no. 2, pp. 127–132, 1997.